

**Medical countermeasures for extraterrestrial environments: Current status and future prospects with focus on acute injuries**

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## **Abstract**

In all space exploratory activities involving humans there are associated risks to health and well-being. Exposure to ionizing radiation represents one of the more serious health concerns. Although medical issues can arise shortly following the rare acute exposures, the more likely radiation-associated injuries will manifest in a delayed fashion following sub-acute exposures or protracted exposures to relatively low radiation doses. The radiological conditions that present these health risks during extraterrestrial space travel are presented in this article, along with current physical and medical countermeasures for such exposure contingencies. The aim of this article is to discuss radiation medical countermeasures that may be considered for future space exploration and travel. Biomedical advances have occurred toward the control and minimization of acute, sub-acute, and fractionated radiation exposure injuries, whether they stem from intentional application of radio-therapeutic procedures or unintentional, accidental terrestrial-based exposures. Those advances, based largely on novel bioengineering, genetic, and combinatorial pharmaceutical strategies, are briefly reviewed here, along with the promising prospects of additional, new and improved medical countering systems/agents that will be forthcoming with additional research and development efforts.

**Key words:** Acute/chronic radiation injuries; galactic cosmic rays; medical countermeasures; space radiation; solar particles; therapeutics

## **Introduction**

The health risks associated with space travel are enormous, but largely manageable with proper assessments and suitable implementation of technologies designed to reduce those risks [1-3]. Considering the National Aeronautics and Space Administration's (NASA's) ambitious future mission plans (e.g., return to the lunar surface with the establishment of a semi-permanent outpost, and possible missions deep into the solar system, such as to the red planet, Mars) and the hazards they present, the agency's Human Research Program has analyzed and prioritized over thirty such unique health risks in terms of mission readiness. These risks need to be evaluated by specific analytics and to find the means to mitigate and control those risks (or simply accept those risks) [1, 2]. Three of the highest level risks, as per estimated rate of occurrence and greatest impact on health and performance during the mission and/or afterward, include that are impacted by space radiation exposure, to varying degrees, include: (1) carcinogenesis; (2) cardiovascular disease; (3) behavioral health and performance decrements [4].

### ***Radiobiological considerations***

Ionizing radiation (IR) exposures of different qualities and of sufficient doses and intensities can cause serious injury to vital cells and tissues of the exposed individual. These injuries will be expressed either early or in a delayed fashion following IR exposure, and will manifest as functionally disruptive structural, physiological, or genetic changes within select organs or organ systems of the body; i.e., pathologic processes within given organs [5, 6]. IR is indiscriminate in terms of targeting of essential biomolecules; this is due to IR's very basic nature of being highly energetic atomic particles/waves that can ionize, and in turn damage, essential cellular components either through direct or indirect means. The indirect route dominates in terms of injury induction and involves the splitting of cellular water by IR (low

linear energy transfer (LET) type such as gamma or X-rays) and the production of free hydroxyl radicals that in turn attack and damage vital cellular constituents [5, 7, 8]. It has been estimated that the major fraction (e.g., two thirds or greater) of all single-strand DNA damage is attributable to hydroxyl radical attack following radiation exposure [7]. High LET type radiation exposures (e.g., heavy ions/particles) provide the exception to the latter, in terms of a direct attack of ionization and damage to vital cellular components [5]. Extraterrestrial space environments with high LET type radiation exposures might pose significantly greater radiological risks with significantly greater health impacts than would Earth-bound terrestrial environments [2].

It has been estimated that astronauts subjected to a major solar particle event (SPE) while traveling within an aluminum ( $10 \text{ g/cm}^2$ ) space vehicle outside a low earth orbit would be exposed to sizable IR doses, in the range of  $\sim 0.07 \text{ Gy-Eq}$  and  $0.2 \text{ Gy}$  to hematopoietic and cutaneous tissues, respectively [9]. However, if the astronauts were in the process of conducting extravehicular work during such a massive SPE event, the estimated radiation doses for these two vital organ systems would be much higher and clearly more hazardous (i.e., IR doses between  $1.38 \text{ Gy-Eq}$  and  $28 \text{ Gy}$  estimated for these organ systems), worst case scenario with doses acquired over 63 hours [9]. Planned extended space missions (i.e., interplanetary travel) will result in sizable cumulative exposures (e.g.,  $1.50\text{-}0.55 \text{ mSv/day}$ ) even under normal conditions of space-weather [10,11]. More specifically, cumulative GCR doses have been estimated at  $950\text{-}1256 \text{ mSv}$  for Mars design reference missions [11]. For beyond low earth orbit, minimal contribution from SPE to cumulative dose are expected due to shielding requirements [12].

Based on prevailing experimental and clinical evidence, chronic exposures to fluences of high LET space particles will pose significant health risks as well; in particular, those risks extend to vision, cognition and behavioral problems, along with hematologic, cardiovascular, pulmonary, gastrointestinal and urogenital syndromes [1, 13-19]. Late-arising cancers represent another major concern [20, 21].

Radiological exposure conditions of major concern within the extraterrestrial space environment certainly include, but are not limited to: (i) solar flares of varying intensities, but all uniformly rich in highly energetic protons, a type of IR generally considered to be a fairly low LET IR; and (ii) protracted streams/fluxes of galactic cosmic radiation (GCR) that are high LET heavy ions and particles [2, 22]. Further, as GCR interacts with metallic sheathing of the space craft, additional IR sources or showers of generally lower energy subatomic particles (neutrons, protons, electrons) are generated, which need to be considered and managed as well [23, 24]. Additionally, GCR or other similar high LET particles pass through soft bodily tissues and progressively tend to lose energy via ionization over the distance traversed, with lower energy spurs, or delta rays, forming penumbras of energetic particles/waves that are subsequently absorbed by tissues [25]. In sum, these radiological conditions associated with extraterrestrial environments present a fundamental problem in terms of radiological protection and need to be considered. The basic mantra of the terrestrially-based health physicist is based on three fundamentally important actions, the three pillars of radiation protection: minimize the time of radiation exposure, maximize the distance between the radiation source and the individual, and use shielding whenever possible. Clearly in the extraterrestrial setting in space, these basic strategies become problematic. First, the astronaut cannot effectively distance within the confines of the space craft; second, time of exposure is generally not adjustable by actions taken

by the astronaut, but rather fixed by the space environment; and third, effective shielding is possible, but limited in terms of basic vehicle design and the use of specially constructed, radioprotective ‘shelters’ (space radiation storm shelters) within the confines of the space vehicle that would be used specifically during intense solar particle flares and fluxes [26]. In general, the best of those shields will be able to block only a limited spectrum of IR. Aboard the space station, the use of hydrogen-rich shielding such as polyethylene in the most frequently occupied locations, such as the sleeping quarters and the galley, has reduced the crew's exposure to space radiation.

The application of safe and effective medical countermeasures for the radiological hazards of the extraterrestrial space environment might be considered to be the fourth pillar of space-associated radiation hygiene. However, this pillar still remains to be fully researched, developed and implemented, along with post-deployment testing and evaluation of effectiveness. These health hazards most prominently include, but are not limited to acute SPE-associated radiation injuries of the skin and of the major vital, internal organ systems of the body (e.g., blood forming and cutaneous systems) and delayed pathologies associated with chronic GCR-type exposures, e.g., central nervous system disturbances (cognitive deficits, motor functions, etc.) and cancer [1, 19, 26-30]

Active coordination between NASA and the United States Food and Drug Administration (FDA) in the development and authorization of space radiation medical countermeasures is required. The application of the FDA’s Animal Rule for countermeasure development and authorization applies to the extraterrestrial space environment [31]. In brief, the Animal Rule was issued by the FDA in 2002 to expedite the development of new drugs and biologics as medical countermeasures against chemical, biological, radiological, and nuclear (CBRN) threats.

This rule applies only to new countermeasures for which conclusive human efficacy investigations under phase II and III clinical trials cannot be performed due to ethical reasons. According to this rule, the FDA can approve new drugs that have been shown to be safe in humans under phase I and effective based on well-controlled animal efficacy studies.

### ***Current status***

Physical countermeasures for extraterrestrial space environments are limited relative to those that are terrestrially-based; examples include specific space craft shielding and sheltering devices, as well as in-flight dosimetry and early warning systems for solar flares and associated SPEs, as well as significant fluences of GCR [26]. In-flight radiation exposure-specific medical countermeasures are limited as well, relative to both the number and scope of effective medicinals. As a consequence, the limited nature of the on-board pharmacy has a potential negative impact on space missions, especially those of long duration and that are outside of the low-Earth orbit. Clearly, mission success hinges on the health and performance of the astronaut crew and, in turn, on having a safe and effective onboard pharmacy [32]. However, improving the ‘spacecraft pharmacy’ is not without challenges, whether they be scientific, logistically or regulatory by nature. Critically important inflight research studies concerning drug pharmacology (along with efficacy, safety and stability) of those drugs of interest are often limited and require the use and extrapolation of data from terrestrial-based work [32].

According to a recent report, the radiation countermeasure component of the on-board emergency medical kit consists of five basic items (Table 1) and all are designed to clinically manage the acute effects of intense SPE exposures (i.e., solar flares) [26]. There are, however, no fully approved medicinals within NASA’s space medical kit designed to specifically counter late-arising pathologies associated with chronic exposure to GCR during prolonged space travel.

Agents intended to be used to clinically manage (and/or prevent) acute SPE exposure-related syndromes are as follows: 1) anti-emetic agents, ondansetron (Zofran) and dexamethasone (Decadron); 2) anti-diarrheal agent, Imodium; 3) antimicrobials/anti-infectives, ciprofloxacin, cephalosporins penicillin, and macroclides; 4) skin topicals for burns/blistering, silver sulfadiazine, corticosteroid creams, topical crystalloid solution, sterile gauze and opioid analgesics; and 5) fluids to manage dehydration. [Note: all these medicinals have been evaluated previously, not only for stability, but also for storability over time [32]. Incorporating a sixth type of medicinal, or more precisely a group of recombinant hematopoietic growth factors (GFs), is currently under consideration for inclusion into the medical emergency kit as well. This group currently consists of the recombinant granulocyte colony-stimulating factor (G-CSF, Neupogen), PEGylated G-CSF (Neulasta), and granulocyte-macrophage colony-stimulating factor (GM-CSF, Leukine), and all are designed to mitigate potentially severe, life-threatening hematopathologic effects (i.e., hematopoietic component of the acute radiation syndrome, H-ARS) of intense SPE exposures during space flight [29, 30, 33-35]. A fourth recombinant GF, Nplate (romiplostim), has been recently approved by the FDA for comparable indications in treating individuals (Earth-bound patients) suffering from acute and intense, unwanted radiation exposures that are at high risk of developing severe thrombocytopenia (suppressed blood platelet levels), uncontrolled bleeding, and consequent fatal outcomes (Table 2) [36]. The clinical use of these recombinants requires prior understanding of an astronaut's hematological status, especially in the days following acute SPE exposure events: Does the blood profile of the astronaut warrant the use of these agents, i.e., is the individual sufficiently granulocytopenic or thrombocytopenic to justify medical intervention? If not, simple follow-up monitoring using on-board clinical assessment tools is essential (automated blood cell counting and cell differential devices). (Note: the

advanced preclinical large animal (nonhuman primate – NHP) studies used to evaluate the efficacy of various recombinant GFs have been conducted largely by using male animals; only in the rhuGM-CSF/Leukine studies has there been a relatively equal distribution of male and female animals employed.)

As a precautionary measure against solar flares and intense SPE exposures, an additional radioprotective agent, namely amifostine, has been carried by astronauts previously during select space (moon) missions; however, as it turned out, the radioprotector was not needed, hence not used. (*Note: The nature and utility of this agent or like agents is discussed later in the manuscript.*)

### ***General strategies for developing and improving space associated radiation medical countermeasures***

One of the major health hazards of deep space travel is the risk of excessive radiation exposures that occur both acutely and chronically. Intense solar flares and associated SPE largely contribute to the risk of acute radiation injury, whereas the ubiquitous, more chronic GCR exposure during space travel is the main concern relative to late-arising or delayed-type injuries. As indicated, basic strategies and associated current medical protocols for countering SPE-associated acute injuries have been developed and implemented, but rely heavily on post-exposure treatments, rather than on the use of preventive or mitigative medicinals. An improved medical strategy might incorporate the selective use of prophylactic/preventive agents, in combination not only with post-exposure injury mitigators, but perhaps therapeutics as well. A polypharmaceutical approach might yield significant improvements, especially if the approach is oriented toward injury prevention and bolstering native radioresistance of the individual astronaut; e.g., the selective use of synergizing combinations of radioprotective pharmaceuticals

and nutraceuticals (Figure 1) [37]. The latter approach to injury prevention would likely serve not only as an effective strategy in medically countering solar flare-associated SPE exposures, but also chronic GCR exposures and related late-arising injuries as well.

### ***Future prospects for improving space radiation related medical countermeasures***

Simply stated, the future looks promising in terms of developing and deploying safe and effective medical countermeasures for future space travelers subjected to potentially hazardous space radiation. Both high tech (newly designed, bioengineered or chemically engineered pharmaceuticals), as well as simpler, more conventional approaches (e.g., use of repurposed medicinals and the use of nutraceuticals) will no doubt come into play (Figure 1) [38]. Regardless, due to the current limited number of such medical radiation countermeasures for extended space travel, coupled with the significant recent scientific, terrestrial-based biomedical advances, there is every expectation that new and improved medicinals for the space traveler will be forthcoming. Nevertheless, there is no doubt that significant challenges lay ahead in getting such improved medicinals into the medical kits of space crafts for future exploratory space missions. Such challenges rest not only on the uncertainties surrounding the impact of various unique stressors (e.g., microgravity) of the space environment on basic drug pharmacology, storage and stability issues, but also on probable changes in the astronaut's basic health and nutritional status that would occur over a prolonged period of time in space.

### ***High tech bio- and chemical engineering approaches***

*Basic discovery of new radioprotective agents via large scale production and screening technologies.* This approach is based on the effective utilization of advanced synthetic and analytic systems for the production and subsequent screening of large arrays of chemical/biochemical/biologic agents with potential, radiation exposure/injury-countering

activities. In general, this approach is actively utilized by biomedical researchers, as well as by the pharmaceutical industry at large; however, in general, it has been underutilized by the radiation biology research community, but it is an approach that clearly needs to be exploited [39, 40].

Specifically, this approach is defined by the combined use of chemical, pharmaceutical and natural product libraries coupled to high throughput screening systems in order to identify brand new, novel chemical or biologic agents that have molecular or cellular targets with defined functionalities. These synthetic/analytic systems have been shown to have significant flexibility and can accommodate a wide variety of research needs and interests; e.g., the potential to conduct phenotypic screening, drug repurposing, and matrix combination screenings of appropriate libraries [41].

*Chemical engineering of native or synthetic radioprotective agents.* It is true that “new medicines require new molecules” and that such novel molecules often come from direct chemical synthesis that follows rationally designed procedures that are augmented by artificial intelligence (AI) by the chemical engineer, which can employ a basic set of chemical engineering tools/principles such as chemical reaction kinetics, thermodynamics, fluid mechanics, and heat and mass transfer [42-44]. The construction of the chemical library is a key component of this new molecule/new medicine discovery process [45]. The radiobiology researcher interested in searching for novel radioprotectants and/or radiomitigators need not ‘reinvent-the-wheel,’ for chemical libraries that are currently available within both the public and private sectors for large scale, high throughput screening for agents having specific functionalities/phenotypes (e.g., specific radioprotective characteristics).

The application of large scale screening of available chemical libraries, however, is not the only viable approach in developing useful medical countermeasures for radiation exposures. The chemical or genetic reengineering of previously well-recognized, specific radioprotective or radiomitigative agents, such as genistein (a natural soybean derived phytoestrogenic moiety), bacterial flagellin (a highly immunogenic, proteinaceous molecule), or novel sulfone-class chemical agents, have all proven to be suitable substrates for such reengineering of select natural products into tentatively useful radioprotective medicinals [46-53]. A sampling of reengineered radioprotective drugs that might be useful for use within the extraterrestrial space environment is listed in Table 3.

*BIO 300, a novel formulation of genistein:* Genistein, 4',5,7-trihydroxyflavone, has demonstrated radioprotective efficacy and is remarkably well-tolerated, as demonstrated in numerous preclinical studies using both small and large animal models of ARS associated injuries [50, 52, 54-57]. [Note: Although 'age' and 'gender' matching processes have been applied in some of these studies, the majority have utilized solely male animals of a standard age].

Under select conditions of prophylaxis, namely subcutaneous (*sc*) injection of genistein over a wide range of doses (25 - 400 mg/kg, ~24 h prior to exposure) provided significant levels of radioprotection (relative to vehicle-treated controls) to subsequent lethal doses of whole-body gamma irradiation in CD2F1 male mice [50, 57]. Furthermore, genistein prophylaxis was exceedingly well tolerated at doses as high as 400 mg/kg, as evidenced by assessments of locomotor activity, grip strength and motor coordination, as well as by body weight, testes weight, and histopathology [57]. Nevertheless, genistein has proven to be challenging from both manufacturing and practical-use perspectives; this is largely due to its low water solubility and

its limitation as primarily an injectable. The preferred oral route was not practicable due to the agent's relatively poor oral bioavailability. The use of synthetic processes (i.e., wet-nanomilling) of genistein to generate nanoparticles has yielded a new genistein formulation (BIO 300) that has significantly improved bioavailability while still retaining significant radioprotective activities to various organ systems regardless of the route of administration [50, 52, 54]. The new BIO 300 product when administered *sc* as single doses 24 – 12 h before total-body irradiation (TBI, 9.25 Gy  $^{60}\text{Co}$   $\gamma$ -radiation), significantly improved 30-d survival in mice [50, 52]. Further, single doses delivered by intramuscular (*im*) injections were also effective when administered up to 2 d prior to acute, lethal radiation exposures. However, BIO 300's optimal dosing regimen was found to be a dose of 150 mg/kg administered 24 h prior to radiation exposure, resulting in a calculated dose reduction factor (DRF) of 1.16 [50]. However, unlike the single *im* injection, optimal oral administrations required twice a day dosing for multiple days prior to radiation exposure in order to demonstrate significant, comparable levels of radioprotection achieved by BIO 300 injection. Interestingly, the level of BIO 300-mediated prophylactic radioprotection is comparable to that seen with a single *sc* injection of the recombinant Neulasta administered after TBI [52].

Prophylaxis with the BIO 300 nanosuspension appears to attenuate irradiation-associated induction of a proinflammatory cytokine storm; namely, the attenuation of interleukin-1 $\beta$  (IL-1 $\beta$ ), IL-6, and cyclooxygenase-2 (COX-2) in mouse bone marrow and spleen. The latter may well serve to protect vital stem and progenitor cell populations within hematopoietic tissues, augmenting subsequent recovery of blood neutrophil and platelet levels [50, 52].

It is important to note that genistein has been shown to decrease adverse effects of radiotherapy and chemotherapy in clinical trials with cancer patients [46]. The oral form of BIO 300 is currently being evaluated in cancer patients undergoing chemotherapy (carboplatin and

paclitaxel) and radiotherapy (radiation exposure of 1.8 – 2 Gy fractions for a total of 60 -70 Gy) for non-small cell lung cancer (NSCLC). Patients received BIO 300 oral formulation daily at doses of 500 mg, 1,000 mg, or 1,500 mg for up to eight weeks. This study is being conducted at multiple medical centers [46]. The oral form of BIO 300 is currently also being evaluated in a phase 2 trial in COVID-19 patients recently discharged from the hospital to determine its effectiveness to mitigate long-term pulmonary injury. In both oncology and COVID-19, the drug's ability to attenuate the inflammatory response resulting from radiation exposure or viral infection is central to therapeutic potential [46].

*Bacterial flagellin and its reengineered analog entolimod (CBLB502).* The second example of a successfully reengineered, natural product is entolimod, a laboratory modified protein extracted from the flagella of select bacterial species. Similar to native flagellin of *Salmonella* species, entolimod has been shown to elicit strong radioprotective effects by stimulating natural killer cells and T lymphocytes through interactions with specific Toll ligand receptor-5 (TLR5) and the subsequent activation of nuclear factors-  $\kappa$ B (NF- $\kappa$ B) [51, 58-60].

Although native flagellin is less than ideal as a potential radioprotectant due to strong immunogenicity, a more useful product was produced by selective chemical/genetic engineering. A truncated form of the flagellin protein with markedly improved characteristics was synthesized, researched and developed as a potential radiation countermeasure [51, 60]. Relative to the lab-engineered protein, the stability and radioprotectiveness of the native protein was retained, while immunogenicity of the protein was substantially reduced. Since its development, entolimod demonstrated significant potential as a medical countermeasure, especially in terms of its capacity to counter the development of two major ARS sub-syndromes, H-ARS and GI-ARS, in murine and NHP models [51, 58-60]. [Note: In early radioprotection studies, female NIH

Swiss/ICR mice were utilized, whereas both male and female NHPs were used for the large animal studies]. However, the lingering immunogenicity of the agent has proven to be problematic, as evidenced by the results of an initial, Phase 1 clinical trial of entolimod's safety and efficacy [61]. Entolimod was found to induce immune response due to memory from earlier exposure to flagellated Enterobacteria [61]. Subsequent development of this agent, however, provides testimony to the utility of current genetic engineering and synthetic biochemical technologies in the research and development (R&D) of ARS countermeasures. A structure-guided reengineering of entolimod was pursued and successfully yielded the next-generation variant, GP532, that was substantially deimmunized, but that retained capacity to induce TLR5-dependent NF- $\kappa$ B activation. The GP532 variant is smaller than the parent agent and has selective mutations that effectively eliminate B and T-cell epitopes and an inflammasome-activating domain [61]. The variant is resistant to human entolimod-neutralizing antibodies, with improved bioavailability and a longer-lasting effect on NF- $\kappa$ B. Most importantly, the new variant, much like the parent, retains potent prophylactic and therapeutic efficacies for irradiation-induced injuries (as per results of mouse model study) [61]. The recent report now suggests that the new and improved entolimod variant has been optimized as a TLR5 agonist and is suitable for patients with high titers of pre-existing flagellin antibodies. Nevertheless, additional assessments of the efficacy and safety of the new entolimod variant is not only warranted, but essential in order to move this agent forward in the process of securing FDA approval for use in treating and managing ARS. In this regard, it should be noted that at least one other 'entolimod-like' agent (KMRCO11) has been bioengineered and fully tested for its radioprotective attributes [62-64].

Biochemical engineering using recombinant DNA technologies. Recombinant DNA technologies permit the novel synthesis of a variety of biologics from living cells (commonly yeast, bacteria, or mammalian cells) that have been artificially redesigned to produce agents not naturally produced by those cells. The technology is now decades old, having initially produced a number of seminal pharmaceuticals and industrial products for global markets [65]. As in the above mentioned situations, the use of these cutting edge technologies has been quite limited in terms of developing novel medicinals for the specific use of countering IR exposures and associated radiation injuries, regardless of whether those radiation exposures are terrestrially or extra-terrestrially based. Nevertheless, the application of these technologies has yielded a number of widely used recombinant GFs that have proven useful in treating a variety of blood disorders (e.g., cancer treatment-induced acute cytopenias) and with a select few have repurposed for treating individuals exposed acutely to unwanted IR (i.e., accidental IR exposures, etc.) who are suffering from H-ARS. This group of recombinants include filgrastim (rhuG-CSF, neupogen), PEGfilgrastim (rhuPEG-CSF, neulasta), sargramostim (rhuGM-CSF, leukine), and romiplastim (rhuTPO, Nplate) and all have been authorized for use by the FDA for H-ARS (as radiomitigators) [40, 66-75]. Additional details on these agents are provided below and in Table 2.

Molecular engineering and the production of chimeric molecules. As pointed out in a previous report “Growth factors and cytokines are integral to tissue microenvironments and the important roles they play in both health and disease, especially in terms of modulating pathogenic processes [40].” They are key components associated with infection, inflammation, and immunity and are double-edged in terms of their capacity to help or to harm the individual. Reengineered (both genetically and chemically) forms of GFs/cytokines have been developed

and investigated over many decades for their therapeutic potentials, including the potential to mitigate and treat acute radiation injuries. New types of recombinant GFs/cytokines with extended therapeutic potentials have been produced; one of these new types are chimeric molecules and are commonly referred to as superkines [76]. The following two chimeric recombinants, namely myelopoietin (MPO) and stable chimeric fibroblast GF (FGF-C), have been tested in a preclinical research setting for their capacity to mitigate acute, potentially fatal radiation injuries in NHPs and rodents, respectively. MPO is a genetically engineered chimeric agent which binds to the G-CSF and IL-3 receptors on hematopoietic cells. The chimeric recombinant has been assessed for its capacity to mitigate potentially fatal hematopoietic injury of male rhesus macaques following acute, whole body  $\gamma$ -irradiation (either 7 or 6 Gy of gamma rayGy). MPO treatments limited both the depth and the duration of the acute irradiation-induced neutropenia and thrombocytopenia, as well as apparently sparing (and promoting recovery) vital bone marrow compartments (e.g., multilineage progenitors) [77-79].

Beyond the design and development of chimeric GFs that solely target hematopoietic tissues, other types of chimeric GFs for other ARS sub-syndromes (e.g., gastrointestinal sub-syndrome or GI-ARS) have been developed and proven to be promisingly reparative in terms of irradiation-associated gastrointestinal tissue injuries. For example, FGF-C is one of such chimerics that is formed by the union of FGF-1 and FGF-2, which strongly stimulates epithelial cell proliferation and, consequently, is endowed with significant reparative activity. When the FGF-C is administered either prophylactically or during the early period following radiation exposure, the agent appears to be efficacious (rodent model of radiation injury) in terms of both protecting and/or repairing epithelia when administered either prophylactically or in the early period following radiation exposure. FGF-C treatments appeared to limit irradiation-induced

apoptosis (cell death), promote crypt cell cycling and the depth of crypt cells, and increase epithelial differentiation in general [80].

*Radioprotective gene transfer.* Basic elements of radioprotective gene transfer technologies and platforms have been pursued experimentally for several decades and with some success clearly being achieved [81-99]. The superoxide dismutase (SOD) gene family and its three principal metalloprotein gene products have been a favorite molecular target for attempting to mitigate the adverse, injurious effects of IR-generated superoxide ( $O_2^-$ ) and related reactive oxygen species (ROS) within sensitive, vital cells and tissues of the body.

In an early study by Suresh *et al.*, several neoplastic human cell lines (i.e., erythroleukemic cells - K562 and melanoma cells - A375) were transfected via electroporation with retroviral constructs bearing the human manganese superoxide dismutase (MnSOD) gene in both the sense and anti-sense orientations, and these different genic constructs were able to modulate the expression of radioprotective MnSOD protein within the transfected cells [81]. In yet another in vitro study, pre-neoplastic murine hematopoietic progenitors (32D cl3) gained radioresistance as they overexpressed MnSOD following transfection with a plasmid containing the human MnSOD gene [91].

The capacity of transfected MnSOD to exert increased radioresistance in intact animals was subsequently demonstrated: in studies with experimental female mice (C3H/HeNHsd) receiving either single or fractionated doses of local (head and neck) irradiation that had been treated orally with liposome-encased MnSOD-plasmid preparations fared better than did placebo-treated control animals. Clinical improvements were noted in terms of decreased mucosal ulceration, lessening of weight loss and overall improved survival [96]. Additional gene transfer platforms for the SODs have been developed for several different tissues/organ systems,

including adenoviral and lentiviral vectors for both acute and delayed, late-arising radiation injuries of pulmonary, vascular and cutaneous tissues [92, 95, 100]. These gene transfer platforms have encompassed not only recombinant viral vectors, but also plasmid liposomes, including minicircle/plasmid liposomes, that have used different routes of delivery (e.g., direct injections/infusions, oral administrations using inhalers, etc.) [101]. It has been demonstrated that the effectiveness of these SOD transgenes are limited, both in time and location. Optimal SOD expression, hence radioprotectiveness, requires specific mitochondrial or cytoplasmic cellular locales, while being constrained to a time frame of ~12 – 28 h [88].

Recent strategies to enhance radioprotection of the individual either by direct gene editing (CRISPR/Cas9) or by ‘vaccination’ using gene-specific mRNA/liposomes have been discussed in reports, but not yet tested experimentally [102-104].

***Simpler protective strategies: Employing nutraceuticals and/or repurposed pharmaceuticals***

*Nutraceuticals*. Nutrition and physiology clearly intersect when it comes to the health and well-being of the individual, whether the individual is earth-bound or in outer space. It is well recognized that the extraterrestrial space environment is unique in terms of its microgravity and radiation, and that these natural space conditions can interrupt basic nutritional/physiological interactions (e.g., risk to infections due to IR/nutrition elicited suppression of immune response). The application of select types of nutraceuticals could well serve to mitigate adverse molecular events, physiological, and/or behavioral decrements arising from either acute solar flares or chronic galactic particle exposures [19, 26, 27, 105-109]. Various forms of tocopherols (i.e., alpha, delta and gamma tocopherols), vitamin A (retinyl acetate), isoflavones (genistein), and a variety of natural fruit extracts have been documented experimentally to provide either significantly increased resistance to the acute, potentially lethal effects of TBI or to exert

radioprotective genomic responses within cells of vital bodily tissues [19, 27, 39, 57, 105-108, 110-112]. All of these agents can be easily administered by various delivery routes (as injectables or as oral supplements), are exceedingly well tolerated with fairly broad time windows of effectiveness, and are quite stable and storable over prolonged periods. For example, prophylaxis of male mice (CD2F1) prior to lethal radiation exposures with vitamin E injections (*sc* doses of 400 mg/kg) yielded substantially increased levels of radioprotection, as evidenced by the estimated DRFs of ~1.23 [105, 113]. Other vitamin E family members, in particular the gamma and delta tocotrienols, are strongly anti-oxidative, with significant levels of demonstrated radioprotectiveness within both small and large animal models [112, 114-116]. A relatively recent report of an advanced preclinical assessment study of gamma-tocotrienol (GT3) efficacy was reported which highlighted the protection from potentially lethal radiation injuries in exposed male and female NHPs [117, 118]. For this study, two different doses of GT3 (37.5 and 75 mg/kg) were tested in animals irradiated with lethal and supra-lethal doses. Specifically, these studies demonstrated that GT3 significantly mitigated acute irradiation-induced H-ARS and fostered an enhanced recovery of blood cell counts (i.e., blood neutrophils and platelets). Most importantly, GT3 prophylaxis elicited recovery was largely independent of supportive care or of the need of multiple doses of recombinant GF treatments (i.e., Neupogen) that would normally accompany the application of the ‘standard of care’ treatments [118]. [Note: There was little difference noted in the responses of the male and female animals, but the small number of animals of each gender utilized limited the statistical power of this study].

While intense exposure(s), whether they be single or multiple exposures, or intense solar flares, will result in acute injuries, more chronic exposures to fluences of highly energetic, heavy galactic particles would elevate the risk of developing significant ‘late arising’ pathologies, the

majority of which have serious health and/or behavioral consequences. The latter most certainly includes both malignant and benign cancers of both solid tissues and of blood-forming tissues (e.g., bone marrow and leukemias), and non-cancerous, late-arising pathologies of virtually every organ system of the body; e.g., ocular, vascular, and nervous systems. These space-associated health risks are ill-defined, as are the ways and means to effectively reduce and counter those risks.

The potential adverse health effects associated with space travel clearly are real and problematic, not only in terms of the basic relationships between conditions of irradiation (dose, dose-rate, radiation quality), but also relative to developing and implementing medical countermeasures. For example, possible irradiation-induced damage to the central nervous system that manifests inflight decrements of cognition, reduced motor function, and behavioral changes, all of which may affect performance and human health [28]. There are, however, reports of preclinical studies that might be instructive, in terms of achieving progress on this daunting space-associated problem. Take for example, the studies by Rabin and colleagues documenting the disruptive effects of neural systems and behavioral patterns in experimental rats exposed (whole-body) fluences of highly energetic charged iron particles (e.g., 1.0 Gy of 1 GeV/n  $^{56}\text{Fe}$  from a dedicated port of a gradient synchrotron at Brookhaven National Laboratory) [17, 119]. Experimental male rats (Sprague-Dawley) were initially pre-conditioned (i.e., pre-trained Morris water maze spatial learning/cognitive performance), then exposed to simulated galactic space IR and subsequently re-evaluated by the performance test nine months following initial exposure. Not surprisingly, the radiation-exposed rats were adversely affected in terms of their cognitive/learning performance with performance decrements that were analogous to a premature aging syndrome [17, 119]. However, by contrast, in another report by the same

researchers that focused on a variety of preconditioning behavioral, learning and memory tasks (i.e., behavioral and cognitive endpoints) in comparably irradiated rats (1.5 and 2.0 Gy doses) that were maintained on strongly antioxidant diets (e.g., ~2% blueberry extract) for weeks prior to heavy particle exposures generally fared better than the irradiated controls (i.e., those IR exposed animals not provided antioxidant-rich diets) in the final, delayed post-exposure cognitive/behavioral tests [107]. Although the effectiveness of the diets clearly served to at least mitigate in part the IR exposure-induced deterioration in performance, the degree of mitigation varied as a function of both specific diets tested and the specific endpoint examined. The study also revealed that animals fed antioxidant diets prior to radiation exposure showed reduced incidence of solid tumors (i.e., at ~1 year post-exposure) as compared to the animals fed the control diet. The authors conclude that high-energy particle irradiation can produce not only age-like decrements in cognitive behavior within standard laboratory animals, but it can also increase tumor formation, and that these radiation-elicited late arising syndromes can be mitigated, at least in part, by specific dietary supplements [107]. As the report concludes, there is little doubt that there are comparable health risks for astronauts during long-term space travel beyond the magnetosphere and that specific dietary regimens might prove useful in countering these adverse health effects [26, 27, 105, 106, 108].

*Probiotics.* As defined by well-characterized microbial preparations (e.g., *Lactobacillus acidophilus*, *Lactobacillus rhamnosus*, and other related *Lactobacillus* species) and endowed with the capacity to alter the makeup of microflora of the individuals' gut microbiome, represent a quite different, but nevertheless important type of nutraceutical strategy for the promotion of gastrointestinal health of the individual, independent of age and gender, and regardless of whether the individual is earth-bound or in deep space [120, 121]. Both preclinical and clinical

observations tend to support the contention that probiotics might have significant medicinal value, especially in terms of mitigating irradiation-induced gastrointestinal injury and associated functional impairments (e.g., reversing the IR effects of microbial dysbiosis, anti-oxidant depletion, loss of intestinal crypt/stem cells and overall mucosal wall integrity, diarrhea, etc.) [122-125]. The underlying basis of probiotics' radioprotective and mitigative actions appear complex, and need to be better characterized. Nevertheless, work in this area continues to yield interesting, informative results; e.g., a rationally designed probiotic, *L. rhamnosus* GG (LGG), is radioprotective when administered via gavage to experimental mice subjected to acute intestinal irradiation and that this radioprotection is "TLR2 and COX-2 dependent and is associated with the migration of COX-2<sup>+</sup> mesenchymal stem cells (MSCs) from the lamina propria of the villus to the lamina propria near the crypt epithelial stem cells" [126]. Further, the authors suggest that the probiotic treatment orchestrates processes involving a timed release of the radioprotective TLR2 agonist, lipoteichoic acid (LTA), followed by a priming of epithelial stem cells and the subsequent triggering of a 'multicellular adaptive immune signaling cascade' [126].

Still another, quite different strategy employed immune gene (IL-22) transgenes being established in gut-associated microbial vectors (e.g., *Lactobacillus reuteri* or *Escherichia coli*) and orally delivered as a dietary probiotic to experimental mice, with the overall intent to bioengineer a more radiation-resilient microbiome of gastrointestines of experimental mice [127]. The recombinant rhuIL-22 directly targets intestinal stem cells (ISCs) and augments growth and expansion of both mouse and human ISCs and is, therefore, inherently reparative for radiation-injured intestinal crypt stem cells [128]. In a proof-of-principle study, experimental mice (C57BL/6) received an IL-22-transgene probiotic at 24 h following acute TBI (9.25 Gy), and the number of recombinant IL-22 positive bacteria within the intestine increased

proportionally to the rise in number of G-protein-coupled receptor 5-expressing intestinal stem cells; a relationship that corresponded to a noted increase in survival (30 d) of the irradiated mice [127].

Repurposed medicinals. This basic strategy has been in play regarding space medicine for some time. Several of these candidate drugs are currently under consideration and will most likely be repurposed in the near future for the extraterrestrial space environment, which include not only the FDA approved recombinant hematopoietic GFs (Neupogen, Neulasta, and Leukine) for treating and mitigating the ARS that arise from intense and unwanted radiation exposures, but also latest of the FDA approved reparative recombinants, romiplostim (Table 2). The intent of incorporating these agents into the space craft's medical kit and pharmacy would be to mitigate the risk of developing life-threatening injuries to the blood forming tissues of astronauts following intense solar flare exposures. An additional word or two on romiplostim is warranted, as this drug might well prove to be exceptionally useful as a space radiation countermeasure. This drug is not only reparative in terms of its actions on select compartments of radiation-injured bone marrow (blood platelet producing megakaryocytes), but it also has a documented survival-sparing effect [73-75]. The drug was developed decades ago by Amgen and was indicated for the fairly rare condition, chronic idiopathic thrombocytopenic purpura (ITP). The drug was FDA approved for marketing in 2003. Romiplostim has been demonstrated to be efficacious in ITP patients in relieving thrombocytopenia via the stimulation of growth and development of marrow megakaryocytes and the associated sustained platelet responses [75, 129, 130]. Based on these attributes, romiplostim was developed as a radiomitigator for H-ARS. Efficacy assessments of the drug using preclinical animal models have continued to support the contention that the drug increases survival within potentially lethally irradiated animals by

supporting and promoting hematopoiesis in bone marrow [40, 73, 74]. [Note: Although these preclinical studies clearly demonstrated an overall, gender-independent survival benefit of romiplostim treatments of acutely irradiated NHPs, female animals were reported to be more sensitive than males to acute, high doses of TBI [73, 74, 131]]

Another FDA-approved recombinant that might be considered for repurposing is palifermin (Kepivance), a recombinant GF (rhuKGF) with reparative capacities for epithelial cells of the body (i.e., cells lining the oral cavity, liver hepatocytes, gastrointestinal tissue, pulmonary cells (type II pneumocytes), hair follicle cells, transitional urothelial cells, and keratinocytes in stratified squamous epithelial cells) [132-134]. Much like the natural keratinocyte GF, palifermin functions in protecting epithelial tissues and repairing their injuries [133-136].

An additional recombinant with considerable therapeutic potential that might be considered for repurposing is oprelvekin (Neumega). This agent was originally licensed by the FDA and marketed for the treatment of chemotherapy-induced acute thrombocytopenia [137] and has a long history of clearly, experimentally demonstrated reparative and survival promoting properties following acute irradiation in experimental animals [138-142]. However, as a radioprotectant, clinical transitioning of this agent has proved to be difficult, largely due to the recombinant's toxicity when delivered systemically [142]. Successful repurposing of this agent will require either a reengineering, or a change in the mode of delivery, or perhaps a narrowing of the range of tissue/organ system targets. Regardless, it has already been demonstrated experimentally that a simple reengineering by PEGylating the recombinant works to improve its overall efficacy (i.e., increasing the time-window of effectiveness, enhancing repair of critically radiation-injured hematopoietic function(s), while reducing the incidence of fatal outcomes)

[143]. Similarly, a change in drug delivery (i.e., from an injectable to either an oral or a direct intraluminal administration) has been shown to be beneficial in terms of reducing the recombinants' systemic toxicity, while still retaining reparative efficacy [140, 142]. [Note: Oprelvikin, as well as palifermin, have strong, well-documented reparative effects on injured epithelia of the body, regardless of species, age or gender].

The aminothiols represent still another important class of radioprotective agents that need to be seriously considered for use in extraterrestrial space flights [144]. Amifostine represents the class's arch-type molecule. Despite amifostine's long history as a well-documented radioprotectant that provides significant levels of radioprotection systemically to properly prophylaxed individuals, the drug still has very limited regulatory approval and only for specific clinical conditions [145]. Due to the toxic 'side-effects' (i.e., nausea, vomiting, general gastrointestinal disturbances, etc.) elicited by the drug when delivered at optimally high radioprotective doses, the agent has not received FDA approval for general public use during nuclear/radiological emergencies, or for space missions. There are, however, anecdotal reports that amifostine was carried by astronauts as a safety precaution during early Apollo moon missions. Regardless, the drug has not been approved by the FDA or NASA for such space mission related purposes. In spite of the drug's limitations (medical and regulatory), research efforts continue to make better use of this very powerful radioprotectant, largely by attempting to limit its toxic side effects [39, 145, 146]. One such research effort (preclinical studies performed largely using male mice) has indicated that by simply reducing the dose of amifostine administered below the threshold for these toxic side effects, some level of radioprotection would remain and might be effectively leveraged by subsequent preventive treatments (e.g., post-exposure GFs and cytokines) [19, 39, 145, 147].

Angiotensin converting enzyme inhibitors (ACEi) function by helping to relax veins and arteries and to lower blood pressure. A large number of these agents are commercially available and are widely used to treat hypertension and heart related problems, but there are also other approved indications as well (e.g., prevention of heart attacks, chronic kidney disease, migraines, and scleroderma). Relative to the problem at hand, it is well documented experimentally that ACEi drugs have the potential to mitigate both early and late-arising irradiation-induced pathologies of several organ systems; most prominently late-arising pathologies of the lung and kidneys of heavily irradiated animals, but also early arising hematopoietic syndromes as well [148-150]. For example, captopril given to experimental rats (Wistar) in drinking water (140 – 180 mg/kg/d) following single, high doses (11 – 15 Gy) of thoracic irradiation clearly spares select lung functions and reduces the incidence of pneumonitis while promoting survival. These measures of ACEi's effectiveness in mitigating irradiation-induced lung injury were utilized to calculate the DRF for specific endpoints related to irradiation-induced lung disease. For overall lung associated morbidity, the dose modifying factors (DMF) were estimated to be 1.07 – 1.17, while for pneumonitis associated tachypnea, estimated DMFs ranged from 1.21 – 1.35 [150].

Relative to the irradiation-induced early arising hematopoietic tissue injury, it was shown experimentally (in a standard acute irradiation-based rodent model) that daily low-dose captopril regimens initiated as late as two days following acute irradiation significantly enhanced overall survival, as did higher drug doses [148]. The drug's capacity to promote survival of acutely irradiated rodents was temporal, perhaps causally related to a promoted hematopoietic recovery phase (i.e., of bone marrow cellularity and blood cells counts) ~3 – 4 weeks following irradiation. Furthermore, captopril treatments did not appear to affect irradiation-induced cell cycle arrest genes or the immediate loss of hematopoietic precursors, but it did reduce the

expression of select hematopoietic GFs/cytokines such as erythropoietin (EPO) and G-CSF in blood plasma. Interestingly, this study reported that captopril treatments, even when delayed (i.e., up to ~48 h post irradiation), appeared to mitigate micro-hemorrhages within brains of radiation-exposed mice [148].

The efficacy of captopril to ward off significant radiation injuries of several major organ systems of the body was demonstrated not only in small experimental animals but also in large animal models as well. For example, in one such study with a mini-pig model of H-ARS was employed and demonstrated that captopril treatments (oral administrations 0.96 mg/kg twice a day for 2 weeks) following TBI ( $^{60}\text{Co}$  1.79 Gy, 0.42 – 0.48 Gy/min) improved the chance of survival of the drug-treated animals (i.e., survival rates of 62.5% in the vehicle-treated control animals compared to 87.5% survival in the captopril-treated group) [148, 151]. Additionally, captopril significantly improved the recovery of peripheral blood mononuclear cells, and trended towards improving the recovery of red cells and platelets. Similar to the results found in the small animal study mentioned earlier, captopril significantly reduced irradiation-induced expression of the cytokines EPO and GM-CSF and an acute-phase inflammatory response cytokine serum amyloid protein A. The latter observation was consistent with a significant suppression of irradiation-induced expression of redox stress genes and improved expression of select hematopoietic cytokines [148].

Statins-3-hydroxy-3-methyl-glutaryl-coenzyme A reductase inhibitors are widely prescribed medicinals that are used for the primary purpose of limiting or reducing blood levels of cholesterol, or more specifically low density lipoprotein fraction of cholesterol and in turn, lowering the risk of cardiovascular disease [152]. As a pharmaceutical group, the statins are considered both safe and effective, especially when appropriate changes in diet and exercise are

applied. The statin group is large with numerous members that include: atorvastatin, rosuvastatin, simvastatin, pravastatin, lovastatin, fluvastatin and pitavastatin [153]. Despite the benefits of statin medications, they are not without side effects. The more common of these include: muscle pain (~10% of the population), headaches, dizziness, tiredness, sickness or weakness, sleep problems, and low platelet counts.

Beyond statin's most positive effect in reducing risks of cardiovascular accidents, there are also several additional medical advantages for using statins; these advantages include, but are not limited to, radioprotection of select types of normal tissues, anti-inflammatory and anti-fibrotic effects, along with the added bonus of being anti-neoplastic under certain circumstances [154]. Although, the precise nature of the statins' anti-neoplastic action is uncertain, it is generally believed that the effect is mediated, in part, by the capacity to delay DNA repair process(es) and by the promotion of neoplastic cell death (apoptosis) [155]. It has been suggested that these drug effects are founded on protein prenylation, rather than its better recognized HMG-coenzyme A reductase inhibitor effect(s) on cholesterol metabolism [155].

Regardless of the uncertainties surrounding the mechanisms of statins' anti-cancer effect(s), it is fairly clear from several epidemiological studies that the extended use of statins serves to ward off irradiation-associated cancers. One report indicated that the daily use of statins (i.e.,  $\geq 4$  years) significantly reduced the risk of developing radiotherapy-associated cancer; e.g., the calculated odds ratios dropped from 0.80 (95% CI 0.66 – 0.96) to 0.64 (95% CI 0.44 – 0.93) [156]. Another retrospective radiotherapy patient cohort study found that the overall cancer mortality rate dropped by approximately 15% in statin-medicated patients [157]. In yet another retrospective cohort study, statin use after a diagnosis of esophageal adenocarcinoma and

subsequent radiotherapy was associated with reduced cancer (esophageal cancers) incidence and overall mortality [158].

By contrast to the statins' apparent suppressive effects on irradiation-associated neoplasia, normal, but injured cells and tissues respond to these drugs in quite the opposite fashion; i.e., in a protective way, as evidenced by enhanced DNA double strand break repair and by limiting the injured cell's overall response to genomic damage, while suppressing subsequent, apoptotic pathways [159]. In addition, statins' anti-inflammatory actions also play a role in the noted protective/tissue reparative processes by largely limiting irradiation-associated inflammation and tissue fibrosis. Specifically, statin treatments appear to suppress not only mRNA expression of both pro-inflammatory and pro-fibrotic cytokine genes, but also dampen neutrophil trafficking to sites of tissue injury, as well as endothelial activation and down-stream inflammatory associated thrombotic responses [160-162].

As mentioned above, the therapeutic attributes of statins have been amply demonstrated both in preclinical work involving experimental animals, as well as in patients enrolled in clinical studies. Select statins have been shown to mitigate significant, life-threatening radiation injuries (or other physicochemical toxicants) of several vital organ systems of the body; e.g., pulmonary, cardiovascular, gastrointestinal and hematopoietic systems [141, 163-166]. For example, in thoracic irradiated rodents, post-exposure treatments of lovastatin limited the buildup of irradiation-induced pro-inflammatory cytokines and presumably, in turn, down-stream tissue fibrosis, while daily, post-irradiation treatments with simvastatin attenuated acute radiation injury of both gastrointestinal and bone marrow tissues [165]. These radiation injury-mitigating effects have been causally linked at the molecular level to statins' targeting of a key protooncogene (i.e., Ras-homologous or Rho) involved in the initial molecular signaling of cell

proliferation and in subsequent p53 regulation of the caspase-dependent apoptosis [159]. Reports of statins' efficacy to mitigate radiation injuries have been extended to the protection of gastrointestinal tissues within cancer patients undergoing radiotherapy as well [141, 167].

Considering the totality of the above reports of statin's positive medicinal effects, it has been rightfully pointed out that these medicinals might be ideally suited for use by astronauts during long space missions, especially under the daily assault of space weather (i.e., space radiation) [144]. In addition to the ACE inhibitors and the statins, a number of other commonly used medicinals have been or are currently being considered for repurposing for the space environment. These include, but not limited to the following: nonsteroidal anti-inflammatory drugs (NSAIDs), N-acetylcysteine, melatonin, metformin, calcium channel blockers, pentoxifylline,  $\beta$ -adrenergic receptor blockers and entolimod [144]. A partial list of medicinals that have been considered for repurposing is shown in Table 4.

#### ***Polypharmaceutical approaches/combinations of effective agents***

Sufficiently intense radiation exposures, acquired either acutely or chronically, either terrestrially or extraterrestrially, have the potential to elicit pathologies within various tissues and organ systems of the body; the very definition of ARS embodies this concept by including an entire complex of often interacting pathologies of different organ systems of the body [168]. As such, it is unlikely that any single drug or medicinal will be sufficient to either prevent, mitigate, or to effectively treat all pathologies within the overall disease complex. Most medical researchers who are involved in this area of study are fully cognizant of the situation and accept the fact that such an optimal, single effective drug (i.e., the silver bullet for ARS) is not close at hand. In lieu of this situation, a polypharmaceutical approach to the problem might represent a reasonable and prudent option. In this regard, there have been a significant number of preclinical

studies that have sought to and have demonstrated the utility of this approach [37, 169, 170]. Further, the overall effectiveness of this approach would be substantially improved by the application of AI and the related decisions concerning the best combination of agents for radioprotection purposes. Briefly, several of the more promising of these different drug combinations are briefly described below, while additional combinations are also listed in Table 5.

Combinations of recombinant GFs and cytokines. The R&D of selecting combinations of recombinant GFs/cytokines for the treatment of H-ARS (terrestrially-based or otherwise) is ongoing and needs to continue in order to improve the efficacy of the current treatment regimens for the ARS. A number of reported preclinical studies using both small and large animal models have demonstrated significant life-sparing effects when select drug combinations have been administered over the early period following acute irradiation [73, 74, 171, 172]. For example, a drug combination consisting of EPO, G-CSF, romiplostim, and nandrolone decanoate was shown to remarkably improve the clinical outcome of acutely irradiated animals receiving the combined treatments; i.e., complete hematological recovery coupled with elimination of early irradiation-associated deaths [171]. The therapeutic utility of using select combinations of GFs/cytokines in treating ARS has been demonstrated pre-clinically using large animal models as well. For example, recent reports by Wong and colleagues showed that the administration of a combination of PEGfilgrastim and romiplostim resulted in significantly improved hematological profiles within acutely irradiated NHPs (as compared to control animals) [40, 73-75] (Table 5).

Combinations of radioprotective and radiomitigative agents. One possible combination would be genistein prophylaxis coupled with post-irradiation captopril treatment (Table 5). Enhanced protection from acute irradiation-associated injury has been demonstrated in a small rodent

model of acute radiation injury (i.e., C57BL/6J mice exposed to lethal doses of 8.25 Gy  $^{60}\text{Co}$  gamma-rays TBI) by combining a prophylactic injection of genistein 24 h prior to exposure and then subsequently following radiation exposure treatment with an ACEi (captopril) for 30 days via oral route of delivery (in drinking water). Results showed that the combined treatment yielded a 95% survival, where genistein prophylaxis alone or post-irradiation treatments with captopril provided far lower survivors, namely 72% and 55%, respectively [173]. As the latter report indicates, the enhanced rates of survival with the combined treatment were directly related to a sparing and recovery of irradiation-associated hematopoietic tissue injury; notably, there was a reduction in irradiation-induced anemia and enhanced recovery of blood cell counts and bone marrow cellularity. A noted early recovery of select progenitor compartments within the marrow was also observed. Interestingly, genistein prophylaxis alone or in combination with captopril post-irradiation treatments seemed to protect marrow progenitors from irradiation-induced micronuclei formation, while captopril alone had no such effect. As other reports have indicated, the inclusion of captopril in the treatment regimen appeared to suppress expression of irradiation-induced EPO [50, 173].

A second possibility for a space-useful drug combination would be prophylaxis with one of the more efficacious aminothiols (e.g., amifostine), coupled with post-irradiation recombinant GFs/cytokine treatments. The radioprotective power of this drug coupling process was clearly demonstrated by several founding proof-of-principal studies [174, 175]. In one such study, a large animal (canine) model of acute radiation injury was employed and used to evaluate the effectiveness of a combined drug regimen (amifostine prophylaxis, plus recombinant hematopoietic growth factor (rhuG-CSF) treatments) to promote survival in irradiated animals over a range of near-lethal to supralethal radiation doses [175]. Groups of animals were

prophylaxed separately with either amifostine alone (150 mg/kg/kg, *iv*), treated post-irradiation with the recombinant hematopoietic GF; G-CSF (10 µg/kg, *sc*), or treated with both amifostine and G-CSF. Administration of a standard clinical support also served as an additional variable. Sixty day survival was used as the primary endpoint and was used to calculate LD<sub>50</sub> values and in turn, DRFs for the different treatment groups. The results were remarkable: the estimated LD<sub>50</sub> value for the untreated irradiated controls was 260 cGy, while for the other treatment groups, the values rose to 340 cGy (for clinical support only), 510 cGy (clinical support, plus G-CSF treatment), 607 cGy (amifostine, without clinical support), 790 cGy (amifostine, plus clinical support), and finally ~1150 cGy (for the combined treatment regimen of amifostine, plus G-CSF, plus clinical support). The DRFs for the latter treatment groups were estimated to be 1.0 (control group), 1.3 (clinical support only), 2.0 (G-CSF, plus clinical support), 2.3 (amifostine prophylaxis, without clinical support), 3.0 (amifostine, plus clinical support), and ~3.9 (full treatment regimen: amifostine, G-CSF and clinical support) [175]. Comparable results were reported as well using a small rodent model of acute radiation injury [174].

*[Note: It needs to be pointed out however, that the doses of amifostine used to prophylax the animals in this study were sufficiently high cause of emesis, and therefore inappropriate for consideration for use during space travel. Nevertheless, the study is clearly instructive in terms of demonstrating what might be possible, what might be achievable by use of the 'polypharmaceutical approach' to radioprotection.]*

A listing of additional promising radiation protective agents that might be used in combination for radioprotection is given in Table 5.

### ***Conclusions***

Relative to the current status, the systems, processes, and tools needed for prevention and treatment of extraterrestrial radiation exposures are indeed available, but limited in scope and capacity to manage such health risks. The general strategic approach currently being taken by the various US federal agencies, including NASA, for the development and improvement of radiation countermeasures is multipronged, involving a mix of both basic and applied physical and biomedical research. We have highlighted a number of these approaches in this report, including high tech bio- and chemical engineering approaches, as well as other approaches that are perhaps somewhat less innovative and cutting edge by nature, but exceedingly useful nonetheless. The latter includes approaches that seek to exploit widely used, over-the-counter nutraceuticals, along with the reexamination of select pharmaceuticals that might be repurposed for the astronaut for use during extraterrestrial space voyages. The repurposing strategy has already paid dividends, as evidenced by the current medicinals already contained within the space traveler's medical kit for the express purpose of clinically managing irradiation-associated overexposure (e.g., anti-nausea, anti-diarrhea drugs, broad spectrum antibiotics, topicals and inflammatories for solar burns, etc.). There is every expectation that the recombinant GFs/cytokines, so prevalent in terrestrial-based medicine, will find their way shortly into medical kits of space vehicles as 'repurposed' medicinals. There is little doubt that a number of ACEi drugs, or perhaps the statins, will be following shortly; as they currently are widely dispensed medicinals that are easily delivered (via the oral route), exceedingly well tolerated, and particularly effective in warding off not only cardiovascular problems, but also under select conditions, both acute and chronic radiation injuries of various vital tissues and organ systems that might manifest under extended times within the space environment.

In the same light, the inclusion of radioprotective nutraceuticals might serve as a ‘simple fix’ in attempting to counter and to manage some of the health risks associated with extended space travel and the inherent exposures to potentially hazardous solar/galactic radiations. This is not simply wishful thinking, but based on well-documented preclinical studies that employed both small and large animal models of radiation injury and that were previously cited regarding the radioprotective/radiomitigative attributes of such nutraceuticals as genistein, gamma tocotrienol, etc. Considering the upside, which includes medical benefits to the space traveler and making use of select types or combinations of nutraceuticals: e.g., ease of oral delivery, effectiveness lasting days to weeks, general lack of toxicity and/or substantial side-effects, easy storage, extended shelf life and stability. NASA has already invested heavily into improving the nutritional health of astronauts, and this effort will most certainly continue into the foreseeable future [176-178]. Any additional nutritional research that impacts space-associated radiation protection will prove to be beneficial from a health standpoint. By comparison, very few, if any, of the more conventional radioprotective pharmaceuticals have all of these positive features. However, the real power of the nutraceutical approach comes into play when selected nutraceuticals (e.g., vitamin E family members) with admittedly modest effectiveness, are combined with other, more potent, radioprotective pharmaceuticals (e.g., amino thiols with sustained release formulations): we refer to this combinatorial approach as the ‘polypharmaceutical strategy’ for improving the quality of radiation medical countermeasures for the extraterrestrial space environment.

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## **Figures**

**Figure 1.** Basic strategies for improving the astronaut's medical kit for excessive space radiation exposures

## **Tables**

**Table 1.** Current radiation countering medicinals within space vehicle's emergency medical kit

**Table 2.** Select recombinant growth factors: Possible candidates for inclusion in the space radiation medical emergency kit

**Table 3.** A representative sampling of reengineered radioprotective drugs for possible use within the extraterrestrial space environment

**Table 4.** A partial list of commonly used and marketed pharmaceuticals with documented radioprotective attributes and with potential to be repurposed for the space environment

**Table 5.** A selection of potentially useful drug combinations that might serve as effective radiation countermeasures for the extraterrestrial environment

## References

- [1] Cucinotta FA, Manuel FK, Jones J, Iszard G, Murrey J, Djojonegro B, et al. Space radiation and cataracts in astronauts. *Radiat Res.* 2001;156:460-6.
- [2] National Council on Radiation Protection and Measurements. Acceptability of risk from radiation—application to human space flight symposium proceeding number 3. 1996. Available at: <https://www.osti.gov/servlets/purl/488809> [Last accessed December 15, 2021]
- [3] National Aeronautics and Space Administration. Space Radiation (eBook) Human Research Program. 2017. Available at: [https://www.nasa.gov/sites/default/files/atoms/files/nasa\\_space\\_radiation\\_ebook\\_0.pdf](https://www.nasa.gov/sites/default/files/atoms/files/nasa_space_radiation_ebook_0.pdf) [Last accessed December 18, 2021]
- [4] Patel ZS, Brunstetter TJ, Tarver WJ, Whitmire AM, Zwart SR, Smith SM, et al. Red risks for a journey to the red planet: The highest priority human health risks for a mission to Mars. *NPJ Microgravity.* 2020;6:33.
- [5] BEIR VII. Molecular and cellular responses to ionizing radiation. In: Council NR, editor. *Health Risks from Exposure to Low Levels of Ionizing Radiation.* Washington, DC: The National Academies Press; 2006.
- [6] Stewart FA, Akleyev AV, Hauer-Jensen M, Hendry JH, Kleiman NJ, Macvittie TJ, et al. ICRP publication 118: ICRP statement on tissue reactions and early and late effects of radiation in normal tissues and organs--threshold doses for tissue reactions in a radiation protection context. *Ann ICRP.* 2012;41:1-322.
- [7] Hall EJ, Giaccia AJ. *Radiobiology for the Radiobiologist.* 7th ed. Philadelphia, PA: Lippincott Williams and Wilkins; 2012.
- [8] International Commission on Radiation Protection. Relative biological effectiveness (RBE), quality factor (Q), and radiation weighting factor (w(R)). A report of the International Commission on Radiological Protection. *Ann ICRP.* 2003;33:1-117.
- [9] National Aeronautics and Space Administration. Evidence Report: Risk of Acute Radiation Syndromes due to Solar Particle Events. 2016. Available at: <https://humanresearchroadmap.nasa.gov/evidence/other/Acute.pdf> [Last accessed April 25, 2022]
- [10] Zeitlin C, Hassler DM, Cucinotta FA, Ehresmann B, Wimmer-Schweingruber RF, Brinza DE, et al. Measurements of energetic particle radiation in transit to Mars on the Mars Science Laboratory. *Science.* 2013;340:1080-4.
- [11] Valinia A, Allen JR, Francisco DR, Minow JI, Pellish JA, Vera AH. *Safe Human Expeditions Beyond Low Earth Orbit (LEO).* 2022. Available at: [https://ntrs.nasa.gov/api/citations/20220002905/downloads/NESC-RP-20-01589\\_NASA-TM-20220002905final.pdf](https://ntrs.nasa.gov/api/citations/20220002905/downloads/NESC-RP-20-01589_NASA-TM-20220002905final.pdf) [Last accessed April 25, 2022]

- [12] National Aeronautics and Space Administration. NASA Space Flight Human-System Standard: NASA-STD-3001, Volume 1 Rrevision B: Crew Health, Table 6, . 2022. Available at: <https://standards.nasa.gov/sites/default/files/standards/NASA/B//2022-01-05-NASA-STD-3001-Vol1-Rev-B-Final-Draft-Signature-010522.pdf> [Last accessed April 25, 2022]
- [13] Abayomi OK. Pathogenesis of irradiation-induced cognitive dysfunction. *Acta Oncol.* 1996;35:659-63.
- [14] Andres-Mach M, Rola R, Fike JR. Radiation effects on neural precursor cells in the dentate gyrus. *Cell Tissue Res.* 2008;331:251-62.
- [15] Chang J, Feng W, Wang Y, Luo Y, Allen AR, Koturbash I, et al. Whole-body proton irradiation causes long-term damage to hematopoietic stem cells in mice. *Radiat Res.* 2015;183:240-8.
- [16] Datta K, Suman S, Kallakury BV, Fornace AJ, Jr. Heavy ion radiation exposure triggered higher intestinal tumor frequency and greater beta-catenin activation than gamma radiation in APC(Min/+) mice. *PloS One.* 2013;8:e59295.
- [17] Rabin BM, Buhler LL, Joseph JA, Shukitt-Hale B, Jenkins DG. Effects of exposure to <sup>56</sup>Fe particles or protons on fixed-ratio operant responding in rats. *J Radiat Res.* 2002;43 Suppl:S225-8.
- [18] Tian J, Pecaut MJ, Slater JM, Gridley DS. Spaceflight modulates expression of extracellular matrix, adhesion, and profibrotic molecules in mouse lung. *J Appl Physiol* (1985). 2010;108:162-71.
- [19] Seed TM. Acute effects. *The Health Risks of Extraterrestrial Environments* 2011. Available at: <https://three.jsc.nasa.gov/articles/SeedAcuteEffects.pdf> [Last accessed July 20, 2021]
- [20] Durante M, Cucinotta FA. Heavy ion carcinogenesis and human space exploration. *Nat Rev Cancer.* 2008;8:465-72.
- [21] National Council on Radiation Protection and Measurements. Relative biological effectiveness of radiations of different quality, NCRP Report number 104. 1990. Available at: <https://ncrponline.org/shop/reports/report-no-104-the-relative-biological-effectiveness-of-radiations-of-different-quality-1990/> [Last accessed December 18, 2021]
- [22] Nelson GA. Space radiation and human exposures, a primer. *Radiat Res.* 2016;185:349-58.
- [23] Heilbronn L. Production of neutrons from interactions of GCR-like particles. In: Wilson JW, Miller J, Konradi A, Cucinotta FA, editors. *Shielding Strategies for Human Space Exploration*. Washington, DC: National Aeronautics and Space Administration; 1997. p. 247-59.
- [24] Rudd EM. HZE interactions in biological materials. In: Wilson JW, Miller J, Konradi A, Cucinotta FA, editors. *Shielding Strategies for Human Space Exploration*. Washington, DC: National Aeronautics and Space Administration; 1997. p. 213-33.

- [25] Curtis SB. Fluence rates, delta rays and cell nucleus hit rates from galactic cosmic rays. 2013. Available at: <https://three.jsc.nasa.gov/articles/tracksinspace.pdf> [Last accessed March 1, 2022]
- [26] Carnell LS. Spaceflight medical countermeasures: a strategic approach for mitigating effects from solar particle events. *Int J Radiat Biol.* 2020;97:S125-S31.
- [27] Kennedy AR. Biological effects of space radiation and development of effective countermeasures. *Life Sci Space Res (Amst).* 2014;1:10-43.
- [28] Cucinotta FA. Space radiation risks to the central nervous system. *Life Sci in Space Res.* 2014;2:54-69.
- [29] Romero-Weaver AL, Wan XS, Diffenderfer ES, Lin L, Kennedy AR. Kinetics of neutrophils in mice exposed to radiation and/or granulocyte colony-stimulating factor treatment. *Radiat Res.* 2013;180:177-88.
- [30] Sanzari JK, Krigsfeld GS, Shuman AL, Diener AK, Lin L, Mai W, et al. Effects of a granulocyte colony stimulating factor, Neulasta, in mini pigs exposed to total body proton irradiation. *Life Sci Space Res (Amst).* 2015;5:13-20.
- [31] U.S. Food and Drug Administration. Animal Rule approvals. 2021. Available at: <https://www.fda.gov/drugs/nda-and-bla-approvals/animal-rule-approvals> [Last accessed November 23, 2021]
- [32] Blue RS, Bayuse TM, Daniels VR, Wotring VE, Suresh R, Mulcahy RA, et al. Supplying a Pharmacy for NASA Exploration Spaceflight: Challenges and Current Understanding. 2019. Available at: <https://ntrs.nasa.gov/api/citations/20190034025/downloads/20190034025.pdf> [Last accessed March 1, 2022]
- [33] Amgen Inc. Neupogen (filgrastim) injection for subcutaneous or intravenous use. 2015. Available at: [http://pi.amgen.com/united\\_states/neupogen/neupogen\\_pi\\_hcp\\_english.pdf](http://pi.amgen.com/united_states/neupogen/neupogen_pi_hcp_english.pdf) [Last accessed April 02, 2021]
- [34] Amgen Inc. Neulasta (pegfilgrastim) injection for subcutaneous use. 2015. Available at: [http://pi.amgen.com/united\\_states/neulasta/neulasta\\_pi\\_hcp\\_english.pdf](http://pi.amgen.com/united_states/neulasta/neulasta_pi_hcp_english.pdf) [Last accessed November 19, 2021]
- [35] Sanofi-Aventis U.S. LLC. LEUKINE® (sargramostim) for injection, for subcutaneous or intravenous use. 2018. Available at: [https://www.accessdata.fda.gov/drugsatfda\\_docs/label/2018/103362s5240lbl.pdf?utm\\_campaign=20180329%20MCMi&utm\\_medium=email&utm\\_source=Eloqua](https://www.accessdata.fda.gov/drugsatfda_docs/label/2018/103362s5240lbl.pdf?utm_campaign=20180329%20MCMi&utm_medium=email&utm_source=Eloqua) [Last accessed December 01, 2021]
- [36] Amgen Inc. NPLATE® (romiplostim) for injection, for subcutaneous use. 2021. Available at: [https://www.accessdata.fda.gov/drugsatfda\\_docs/label/2021/125268s1671bl.pdf](https://www.accessdata.fda.gov/drugsatfda_docs/label/2021/125268s1671bl.pdf) [Last accessed December 14, 2021]

- [37] Gasperetti T, Miller T, Gao F, Narayanan J, Jacobs ER, Szabo A, et al. Polypharmacy to mitigate acute and delayed radiation syndromes. *Front Pharmacol.* 2021;12:634477.
- [38] Singh VK, Seed TM. Repurposing pharmaceuticals previously approved by regulatory agencies to medically counter injuries arising either early or late following radiation exposure. *Front Pharmacol.* 2021;12:624844.
- [39] Seed TM. Radiation protectants: current status and future prospects. *Health Phys.* 2005;89:531-45.
- [40] Singh VK, Seed TM. Radiation countermeasures for hematopoietic acute radiation syndrome: growth factors, cytokines and beyond. *Int J Radiat Biol.* 2021;97:1526-47.
- [41] National Center for Advancing Translational Sciences. Assay Development & Screening. 2020. Available at: <https://ncats.nih.gov/preclinical/drugdev/assay> [Last accessed December 16, 2021]
- [42] Freedman DH. Hunting for new drugs with AI. *Nature.* 2019;576:S49-S53.
- [43] National Research Council. Challenges for the Chemical Sciences in the 21st Century. Washington, DC: National Academies Press; 2004.
- [44] Ende am DJ, Ende am MT. *Chemical Engineering in the Pharmaceutical Industry: Active Pharmaceutical Ingredients.* 2nd Edition ed. Hoboken, NJ: John Wiley and Sons; 2019.
- [45] Saldivar-Gonzalez FI, Huerta-Garcia CS, Medina-Franco JL. Chemoinformatics-based enumeration of chemical libraries: a tutorial. *J Cheminform.* 2020;12:64.
- [46] Humanetics Pharmaceuticals. BIO 300 – A unique, highly selective radiation modulator. 2021. Available at: <https://www.humaneticscorp.com/> [Last accessed December 03, 2021]
- [47] Suman S, Datta K, Doiron K, Ren C, Kumar R, Taft DR, et al. Radioprotective effects of ON 01210.Na upon oral administration. *J Radiat Res.* 2012;53:368-76.
- [48] Suman S, Maniar M, Fornace AJ, Jr., Datta K. Administration of ON 01210.Na after exposure to ionizing radiation protects bone marrow cells by attenuating DNA damage response. *Radiat Oncol.* 2012;7:6.
- [49] Singh VK, Seed TM. Pharmacological management of ionizing radiation injuries: Current and prospective agents and targeted organ systems. *Expert Opin Pharmacother.* 2020;21:317-37.
- [50] Singh VK, Seed TM. BIO 300: a promising radiation countermeasure under advanced development for acute radiation syndrome and the delayed effects of acute radiation exposure. *Exp Opin Invest Drugs.* 2020;29:429-41.
- [51] Singh VK, Seed TM. Entolimod as a radiation countermeasure for acute radiation syndrome. *Drug Discov Today.* 2021;26:17-30.

- [52] Singh VK, Fatanmi OO, Wise SY, Carpenter A, Nakamura-Peek S, Serebrenik AA, et al. A novel oral formulation of BIO 300 confers prophylactic radioprotection from acute radiation syndrome in mice. *Int J Radiat Biol.* 2022;(in press).
- [53] Tang L, Peng T, Wang G, Wen X, Sun Y, Zhang S, et al. Synthesis and radioprotective effects of novel benzyl naphthyl sulfoxide (sulfone) derivatives transformed from Ex-RAD. *Medchemcomm.* 2018;9:625-31.
- [54] Jackson IL, Zodda A, Gurung G, Pavlovic R, Kaytor MD, Kuskowski MA, et al. BIO 300, a nanosuspension of genistein, mitigates pneumonitis/fibrosis following high-dose radiation exposure in the C57L/J murine model. *Br J Pharmacol.* 2017;174:4738-50.
- [55] Cheema AK, Mehta KY, Santiago PT, Fatanmi OO, Kaytor MD, Singh VK. Pharmacokinetic and metabolomic studies with BIO 300, a nanosuspension of genistein, in a nonhuman primate model. *Int J Mol Sci.* 2019;20:1231.
- [56] Cheema AK, Li Y, Singh J, Johnson R, Girgis M, Wise SY, et al. Microbiome study in irradiated mice treated with BIO 300, a promising radiation countermeasure. *Anim Microbiome.* 2021;3:71.
- [57] Landauer MR, Srinivasan V, Seed TM. Genistein treatment protects mice from ionizing radiation injury. *J Appl Toxicol.* 2003;23:379-85.
- [58] Krivokrysenko VI, Toshkov IA, Gleiberman AS, Krasnov P, Shyshynova I, Bespalov I, et al. The Toll-like receptor 5 agonist Entolimod mitigates lethal acute radiation syndrome in non-human primates. *PLoS One.* 2015;10:e0135388.
- [59] Krivokrysenko VI, Shakhov AN, Singh VK, Bone F, Kononov Y, Shyshynova I, et al. Identification of granulocyte colony-stimulating factor and interleukin-6 as candidate biomarkers of CBLB502 efficacy as a medical radiation countermeasure. *J Pharmacol Exp Therapt.* 2012;343:497-508.
- [60] Burdelya LG, Krivokrysenko VI, Tallant TC, Strom E, Gleiberman AS, Gupta D, et al. An agonist of Toll-like receptor 5 has radioprotective activity in mouse and primate models. *Science.* 2008;320:226-30.
- [61] Mett V, Kurnasov OV, Bespalov IA, Molodtsov I, M. Brackett CM, Burdelya LG, et al. A deimmunized and pharmacologically optimized Toll-like receptor 5 agonist for therapeutic applications. *Commun Biol.* 2021;4:466.
- [62] Kim JY, Park JH, Seo SM, Park JI, Jeon HY, Lee HK, et al. Radioprotective effect of newly synthesized toll-like receptor 5 agonist, KMRC011, in mice exposed to total-body irradiation. *J Radiat Res.* 2019;60:432-41.
- [63] Song WS, Kim JH, Choi CM, Lee WJ, Yoon SI. TLR5 binding and activation by KMRC011, a flagellin-derived radiation countermeasure. *Biochem Biophys Res Commun.* 2019;508:570-5.

- [64] Lee HS, Cho DW, Han JS, Han SC, Woo SK, Jun SY, et al. KMRC011, an agonist of toll-like receptor 5, mitigates irradiation-induced tissue damage and mortality in cynomolgus monkeys. *J Immunotoxicol.* 2020;17:31-42.
- [65] Stryjewska A, Kiepusa K, Librowski T, Lochynski S. Biotechnology and genetic engineering in the new drug development. Part I. DNA technology and recombinant proteins. *Pharmacol Rep.* 2013;65:1075-85.
- [66] Gale RP, Armitage JO. Use of molecularly-cloned haematopoietic growth factors in persons exposed to acute high-dose, high-dose rate whole-body ionizing radiations. *Blood Rev.* 2020;45:100690.
- [67] Lazarus HM, Armitage JO, Gale RP. Role of molecularly-cloned hematopoietic growth factors after acute high-dose radiation exposures. *J Radiol Prot.* 2021;10:41.
- [68] Farese AM, Cohen MV, Katz BP, Smith CP, Gibbs A, Cohen DM, et al. Filgrastim improves survival in lethally irradiated nonhuman primates. *Radiat Res.* 2013;179:89-100.
- [69] Hankey KG, Farese AM, Blaauw EC, Gibbs AM, Smith CP, Katz BP, et al. Pegfilgrastim improves survival of lethally irradiated nonhuman primates. *Radiat Res.* 2015;183:643-55.
- [70] Singh VK, Seed TM. An update on sargramostim for treatment of acute radiation syndrome. *Drugs Today (Barc).* 2018;54:679-93.
- [71] Clayton NP, Khan-Malek RC, Dangler CA, Zhang D, Ascah A, Gains M, et al. Sargramostim (rhu GM-CSF) improves survival of non-human primates with severe bone marrow suppression after acute, high-dose, whole-body irradiation. *Radiat Res.* 2021;195:191-9.
- [72] Zhong Y, Pouliot M, Downey AM, Mockbee C, Roychowdhury D, Wierzbicki W, et al. Efficacy of delayed administration of sargramostim up to 120 hours post exposure in a nonhuman primate total body radiation model. *Int J Radiat Biol.* 2021;97:S100-S16.
- [73] Wong K, Bunin DI, Bujold K, Javitz HS, Bakke J, Gahagen J, et al. Romiplostim (Nplate) alone and in combination with pegfilgrastim (Neulasta) increased survival and reduces incidence, duration, and severity of thrombocytopenia post-irradiation in non-human primates. Annual Conference of Radiation Research Society. Virtual2020.
- [74] Wong K, Chang PY, Fielden M, Downey AM, Bunin D, Bakke J, et al. Pharmacodynamics of romiplostim alone and in combination with pegfilgrastim on acute radiation-induced thrombocytopenia and neutropenia in non-human primates. *Int J Radiat Biol.* 2020;96:155-66.
- [75] Singh VK, Seed TM. An update on romiplostim for treatment of acute radiation syndrome. *Drugs Today (Barc).* 2022;58:133-45.
- [76] Conlon KC, Miljkovic MD, Waldmann TA. Cytokines in the Treatment of Cancer. *J Interferon Cytokine Res.* 2019;39:6-21.

- [77] MacVittie TJ, Farese AM, Smith WG, Baum CM, Burton E, McKearn JP. Myelopoietin, an engineered chimeric IL-3 and G-CSF receptor agonist, stimulates multilineage hematopoietic recovery in a nonhuman primate model of radiation-induced myelosuppression. *Blood*. 2000;95:837-45.
- [78] Farese AM, Casey DB, Smith WG, Vigneulle RM, McKearn JP, MacVittie TJ. Leridistim, a chimeric dual G-CSF and IL-3 receptor agonist, enhances multilineage hematopoietic recovery in a nonhuman primate model of radiation-induced myelosuppression: effect of schedule, dose, and route of administration. *Stem Cells*. 2001;19:522-33.
- [79] Farese AM, Smith WG, Giri JG, Siegel N, McKearn JP, MacVittie TJ. Promegapoietin-1a, an engineered chimeric IL-3 and Mpl-L receptor agonist, stimulates hematopoietic recovery in conventional and abbreviated schedules following radiation-induced myelosuppression in nonhuman primates. *Stem Cells*. 2001;19:329-38.
- [80] Nakayama F, Hagiwara A, Umeda S, Asada M, Goto M, Oki J, et al. Post treatment with an FGF chimeric growth factor enhances epithelial cell proliferation to improve recovery from radiation-induced intestinal damage. *Int J Radiat Oncol Biol Phys*. 2010;78:860-7.
- [81] Suresh A, Tung F, Moreb J, Zucali JR. Role of manganese superoxide dismutase in radioprotection using gene transfer studies. *Cancer Gene Ther*. 1994;1:85-90.
- [82] Stickle RL, Epperly MW, Klein E, Bray JA, Greenberger JS. Prevention of irradiation-induced esophagitis by plasmid/liposome delivery of the human manganese superoxide dismutase transgene. *Radiat Oncol Investig*. 1999;7:204-17.
- [83] Kanai AJ, Zeidel ML, Lavelle JP, Greenberger JS, Birder LA, de Groat WC, et al. Manganese superoxide dismutase gene therapy protects against irradiation-induced cystitis. *Am J Physiol Renal Physiol*. 2002;283:F1304-12.
- [84] Guo H, Seixas-Silva JA, Jr., Epperly MW, Gretton JE, Shin DM, Bar-Sagi D, et al. Prevention of radiation-induced oral cavity mucositis by plasmid/liposome delivery of the human manganese superoxide dismutase (SOD2) transgene. *Radiat Res*. 2003;159:361-70.
- [85] Carpenter M, Epperly MW, Agarwal A, Nie S, Hricisak L, Niu Y, et al. Inhalation delivery of manganese superoxide dismutase-plasmid/liposomes protects the murine lung from irradiation damage. *Gene Ther*. 2005;12:685-93.
- [86] Rajagopalan MS, Stone B, Rwigema JC, Salimi U, Epperly MW, Goff J, et al. Intraesophageal manganese superoxide dismutase-plasmid liposomes ameliorates novel total-body and thoracic radiation sensitivity of NOS1<sup>-/-</sup> mice. *Radiat Res*. 2010;174:297-312.
- [87] Maier P, Veldwijk MR, Wenz F. Radioprotective gene therapy. *Expert Opin Biol Ther*. 2011;11:1135-51.
- [88] Greenberger JS, Mukherjee A, Epperly MW. Gene therapy for systemic or organ specific delivery of manganese superoxide dismutase. *Antioxidants*. 2021;10.

- [89] Epperly M, Bray J, Kraeger S, Zwacka R, Engelhardt J, Travis E, et al. Prevention of late effects of irradiation lung damage by manganese superoxide dismutase gene therapy. *Gene Ther.* 1998;5:196-208.
- [90] Epperly MW, Bray JA, Carlos TM, Prochownik E, Greenberger JS. Biology of marrow stromal cell lines derived from long-term bone marrow cultures of Trp53-deficient mice. *Radiat Res.* 1999;152:29-40.
- [91] Epperly MW, Bray JA, Esocobar P, Bigbee WL, Watkins S, Greenberger JS. Overexpression of the human manganese superoxide dismutase (MnSOD) transgene in subclones of murine hematopoietic progenitor cell line 32D cl 3 decreases irradiation-induced apoptosis but does not alter G2/M or G1/S phase cell cycle arrest. *Radiat Oncol Investig.* 1999;7:331-42.
- [92] Epperly MW, Bray JA, Krager S, Berry LM, Gooding W, Engelhardt JF, et al. Intratracheal injection of adenovirus containing the human MnSOD transgene protects athymic nude mice from irradiation-induced organizing alveolitis. *Int J Radiat Oncol Biol Phys.* 1999;43:169-81.
- [93] Epperly MW, Travis EL, Sikora C, Greenberger JS. Manganese [correction of Magnesium] superoxide dismutase (MnSOD) plasmid/liposome pulmonary radioprotective gene therapy: modulation of irradiation-induced mRNA for IL-1, TNF-alpha, and TGF-beta correlates with delay of organizing alveolitis/fibrosis. *Biol Blood Marrow Transplant.* 1999;5:204-14.
- [94] Epperly MW, Kagan VE, Sikora CA, Gretton JE, Defilippi SJ, Bar-Sagi D, et al. Manganese superoxide dismutase-plasmid/liposome (MnSOD-PL) administration protects mice from esophagitis associated with fractionated radiation. *Int J Cancer.* 2001;96:221-31.
- [95] Epperly MW, Defilippi S, Sikora C, Gretton J, Greenberger JS. Radioprotection of lung and esophagus by overexpression of the human manganese superoxide dismutase transgene. *Mil Med.* 2002;167:71-3.
- [96] Epperly MW, Carpenter M, Agarwal A, Mitra P, Nie S, Greenberger JS. Intraoral manganese superoxide dismutase-plasmid/liposome (MnSOD-PL) radioprotective gene therapy decreases ionizing irradiation-induced murine mucosal cell cycling and apoptosis. *In Vivo.* 2004;18:401-10.
- [97] Epperly MW, Wegner R, Kanai AJ, Kagan V, Greenberger EE, Nie S, et al. Effects of MnSOD-plasmid liposome gene therapy on antioxidant levels in irradiated murine oral cavity orthotopic tumors. *Radiat Res.* 2007;167:289-97.
- [98] Epperly MW, Smith T, Zhang X, Goff JP, Franicola D, Greenberger B, et al. Modulation of in utero total body irradiation induced newborn mouse growth retardation by maternal manganese superoxide dismutase-plasmid liposome (MnSOD-PL) gene therapy. *Gene Ther.* 2011;18:579-83.
- [99] Epperly MW, Wang H, Jones JA, Dixon T, Montesinos CA, Greenberger JS. Antioxidant-chemoprevention diet ameliorates late effects of total-body irradiation and supplements radioprotection by MnSOD-plasmid liposome administration. *Radiat Res.* 2011;175:759-65.

- [100] Khan AA, Paget JT, McLaughlin M, Kyula JN, Wilkinson MJ, Pencavel T, et al. Genetically modified lentiviruses that preserve microvascular function protect against late radiation damage in normal tissues. *Sci Transl Med*. 2018;10.
- [101] Everett WH, Curiel DT. Gene therapy for radioprotection. *Cancer Gene Ther*. 2015;22:172-80.
- [102] Borg AM, Baker JE. Contemporary biomedical engineering perspective on volitional evolution for human radiotolerance enhancement beyond low-earth orbit. *Synth Biol (Oxf)*. 2021;6:ysab023.
- [103] Maliev V, Popov D, Casey RC, Jones JA. Mechanisms of action for an anti-radiation vaccine in reducing the biological impact of high dose and dose-rate, low-linear energy transfer radiation exposure. *Radiats Biol Radioecol*. 2007;47:286-91.
- [104] Popov D. Radiation protection: Ribonucleic acid (RNA) antiradiation vaccine or messenger RNA (mRNA) antiradiation vaccine. 2021. Available at: [https://www.researchgate.net/publication/352414347\\_Radiation\\_Protection\\_Ribonucleic\\_acid\\_RNA\\_Antiradiation\\_vaccine\\_or\\_messenger\\_RNA\\_mRNA\\_Antiradiation\\_vaccine](https://www.researchgate.net/publication/352414347_Radiation_Protection_Ribonucleic_acid_RNA_Antiradiation_vaccine_or_messenger_RNA_mRNA_Antiradiation_vaccine) [Last accessed December 17, 2021]
- [105] Seed T, Kumar S, Whitnall M, Srinivasan V, Singh V, Elliott T, et al. New strategies for the prevention of radiation injury: possible implications for countering radiation hazards of long-term space travel. *J Radiat Res*. 2002;43 Suppl:S239-44.
- [106] Burns FJ, Chen S, Xu G, Wu F, Tang MS. The action of a dietary retinoid on gene expression and cancer induction in electron-irradiated rat skin. *J Radiat Res*. 2002;43 Suppl:S229-32.
- [107] Rabin BM, Shukitt-Hale B, Joseph J, Todd P. Diet as a factor in behavioral radiation protection following exposure to heavy particles. *Gravit Space Biol Bull*. 2005;18:71-7.
- [108] Montesinos CA, Khalid R, Cristea O, Greenberger JS, Epperly MW, Lemon JA, et al. Space radiation protection countermeasures in microgravity and planetary exploration. *Life (Basel)*. 2021;11.
- [109] Shirazi A, Ghobadi G, Ghazi-Khansari M. A radiobiological review on melatonin: a novel radioprotector. *J Radiat Res*. 2007;48:263-72.
- [110] Weiss JF, Landauer MR. Protection against ionizing radiation by antioxidant nutrients and phytochemicals. *Toxicology*. 2003;189:1-20.
- [111] Lemon JA, Boreham DR, Rollo CD. A dietary supplement abolishes age-related cognitive decline in transgenic mice expressing elevated free radical processes. *Exp Biol Med (Maywood)*. 2003;228:800-10.
- [112] Singh VK, Beattie LA, Seed TM. Vitamin E: Tocopherols and tocotrienols as potential radiation countermeasures. *J Radiat Res*. 2013;54:973-88.

- [113] Kumar KS, Srinivasan V, Toles R, Jobe L, Seed TM. Nutritional approaches to radioprotection: vitamin E. *Mil Med.* 2002;167:57-9.
- [114] Satyamitra M, Ney P, Graves J, 3rd, Mullaney C, Srinivasan V. Mechanism of radioprotection by delta-tocotrienol: pharmacokinetics, pharmacodynamics and modulation of signalling pathways. *Br J Radiol.* 2012;85:e1093-103.
- [115] Ghosh SP, Kulkarni S, Hieber K, Toles R, Romanyukha L, Kao TC, et al. Gamma-tocotrienol, a tocol antioxidant as a potent radioprotector. *Int J Radiat Biol.* 2009;85:598-606.
- [116] Singh VK, Singh PK, Wise SY, Posarac A, Fatanmi OO. Radioprotective properties of tocopherol succinate against ionizing radiation in mice. *J Radiat Res.* 2013;54:210-20.
- [117] Singh VK, Hauer-Jensen M. Gamma-tocotrienol as a promising countermeasure for acute radiation syndrome: Current status. *Int J Mol Sci.* 2016;17:e663.
- [118] Singh VK, Kulkarni S, Fatanmi OO, Wise SY, Newman VL, Romaine PL, et al. Radioprotective efficacy of gamma-tocotrienol in nonhuman primates. *Radiat Res.* 2016;185:285-98.
- [119] Shukitt-Hale B, Casadesus G, Cantuti-Castelvetri I, Rabin BM, Joseph JA. Cognitive deficits induced by <sup>56</sup>Fe radiation exposure. *Adv Space Res.* 2003;31:119-26.
- [120] Segers C, Verslegers M, Baatout S, Leys N, Lebeer S, Mastroleo F. Food supplements to mitigate detrimental effects of pelvic radiotherapy. *Microorganisms.* 2019;7.
- [121] Obrador E, Salvador R, Villaescusa JI, Soriano JM, Estrela JM, Montoro A. Radioprotection and radiomitigation: from the bench to clinical practice. *Biomedicines.* 2020;8.
- [122] Ciorba MA, Stenson WF. Probiotic therapy in radiation-induced intestinal injury and repair. *Ann N Y Acad Sci.* 2009;1165:190-4.
- [123] Devaraj NK, Suppiah S, Veettil SK, Ching SM, Lee KW, Menon RK, et al. The effects of probiotic supplementation on the incidence of diarrhea in cancer patients receiving radiation therapy: a systematic review with meta-analysis and trial sequential analysis of randomized controlled trials. *Nutrients.* 2019;11.
- [124] Demirer S, Aydintug S, Aslim B, Kepenekci I, Sengul N, Evirgen O, et al. Effects of probiotics on radiation-induced intestinal injury in rats. *Nutrition.* 2006;22:179-86.
- [125] Spyropoulos BG, Misiakos EP, Fotiadis C, Stoidis CN. Antioxidant properties of probiotics and their protective effects in the pathogenesis of radiation-induced enteritis and colitis. *Dig Dis Sci.* 2011;56:285-94.
- [126] Riehl TE, Alvarado D, Ee X, Zuckerman A, Foster L, Kapoor V, et al. *Lactobacillus rhamnosus* GG protects the intestinal epithelium from radiation injury through release of lipoteichoic acid, macrophage activation and the migration of mesenchymal stem cells. *Gut.* 2019;68:1003-13.

- [127] Zhang X, Fisher R, Hou W, Shields D, Epperly MW, Wang H, et al. Second-generation probiotics producing IL-22 increase survival of mice after total body irradiation. *In Vivo*. 2020;34:39-50.
- [128] Lindemans CA, Calafiore M, Mertelsmann AM, O'Connor MH, Dudakov JA, Jenq RR, et al. Interleukin-22 promotes intestinal-stem-cell-mediated epithelial regeneration. *Nature*. 2015;528:560-4.
- [129] Parameswaran R, Lunning M, Mantha S, Devlin S, Hamilton A, Schwartz G, et al. Romiplostim for management of chemotherapy-induced thrombocytopenia. *Support Care Cancer*. 2014;22:1217-22.
- [130] Jacobson AE, Shah N, Setty BA. Romiplostim for therapy-related thrombocytopenia in pediatric malignancies. *Pediatr Blood Cancer*. 2017;64:e26473.
- [131] Beach T, Authier S, Javitz HS, Wong K, Bakke J, Gahagen J, et al. Total body irradiation models in NHPs - consideration of animal sex and provision of supportive care to advance model development. *Int J Radiat Biol*. 2020:1-5.
- [132] Farrell CL, Bready JV, Rex KL, Chen JN, DiPalma CR, Whitcomb KL, et al. Keratinocyte growth factor protects mice from chemotherapy and radiation-induced gastrointestinal injury and mortality. *Cancer Res*. 1998;58:933-9.
- [133] Farrell CL, Rex KL, Kaufman SA, Dipalma CR, Chen JN, Scully S, et al. Effects of keratinocyte growth factor in the squamous epithelium of the upper aerodigestive tract of normal and irradiated mice. *Int J Radiat Biol*. 1999;75:609-20.
- [134] Danilenko DM. Preclinical and early clinical development of keratinocyte growth factor, an epithelial-specific tissue growth factor. *Toxicol Pathol*. 1999;27:64-71.
- [135] Finch PW, Mark Cross LJ, McAuley DF, Farrell CL. Palifermin for the protection and regeneration of epithelial tissues following injury: new findings in basic research and pre-clinical models. *J Cell Mol Med*. 2013;17:1065-87.
- [136] Beaven AW, Shea TC. The effect of palifermin on chemotherapy and radiation therapy-induced mucositis: a review of the current literature. *Support Cancer Ther*. 2007;4:188-97.
- [137] Kaye JA. FDA licensure of NEUMEGA to prevent severe chemotherapy-induced thrombocytopenia. *Stem Cells*. 1998;16 Suppl 2:207-23.
- [138] Potten CS. Protection of the small intestinal clonogenic stem cells from radiation-induced damage by pretreatment with interleukin 11 also increases murine survival time. *Stem Cells*. 1996;14:452-9.
- [139] Seed TM, Inal CE, Deen JE. Assessment of a combined G-CSF plus IL-11 cytokine treatment for radiation-induced hematopoietic injury. 48th Annual Meeting of the Radiation Research Society. San Juan, Puerto Rico 2001. p. 161.

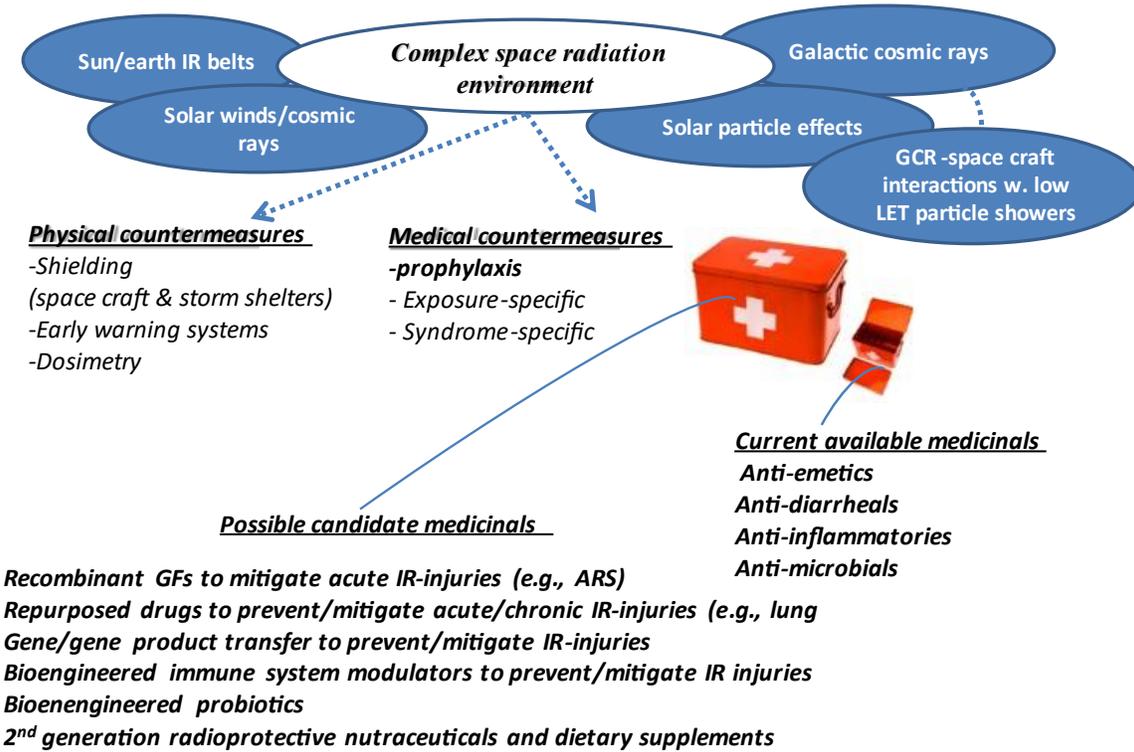
- [140] Boerma M, Wang J, Burnett AF, Santin AD, Roman JJ, Hauer-Jensen M. Local administration of interleukin-11 ameliorates intestinal radiation injury in rats. *Cancer Res.* 2007;67:9501-6.
- [141] Hauer-Jensen M, Wang J, Boerma M, Fu Q, Denham JW. Radiation damage to the gastrointestinal tract: mechanisms, diagnosis, and management. *Curr Opin Support Palliat Care.* 2007;1:23-9.
- [142] Burnett AF, Biju PG, Lui H, Hauer-Jensen M. Oral interleukin 11 as a countermeasure to lethal total-body irradiation in a murine model. *Radiat Res.* 2013;180:595-602.
- [143] Kumar VP, Biswas S, Sharma NK, Stone S, Fam CM, Cox GN, et al. PEGylated IL-11 (BBT-059): A novel radiation countermeasure for hematopoietic acute radiation syndrome. *Health Phys.* 2018;115:65-76.
- [144] McLaughlin MF, Donoviel DB, Jones JA. Novel indications for commonly used medications as radiation protectants in spaceflight. *Aerosp Med Hum Perform.* 2017;88:665-76.
- [145] Singh VK, Seed TM. The efficacy and safety of amifostine for the acute radiation syndrome. *Expert Opin Drug Saf.* 2019;18:1077-90.
- [146] Srinivasan V, Pendergrass JA, Jr., Kumar KS, Landauer MR, Seed TM. Radioprotection, pharmacokinetic and behavioural studies in mouse implanted with biodegradable drug (amifostine) pellets. *Int J Radiat Biol.* 2002;78:535-43.
- [147] Hofer M, Hoferova Z, Depes D, Falk M. Combining pharmacological countermeasures to attenuate the acute radiation syndrome -A concise review. *Molecules.* 2017;22:834.
- [148] McCart EA, Lee YH, Jha J, Mungunsukh O, Rittase WB, Summers TA, Jr., et al. Delayed captopril administration mitigates hematopoietic injury in a murine model of total body irradiation. *Sci Rep.* 2019;9:2198.
- [149] Moulder JE, Cohen EP, Fish BL. Mitigation of experimental radiation nephropathy by renin-equivalent doses of angiotensin converting enzyme inhibitors. *Int J Radiat Biol.* 2014;90:762-8.
- [150] Medhora M, Gao F, Jacobs ER, Moulder JE. Radiation damage to the lung: mitigation by angiotensin-converting enzyme (ACE) inhibitors. *Respirology.* 2012;17:66-71.
- [151] Rittase WB, McCart EA, Muir JM, Bouten RM, Slaven JE, Mungunsukh O, et al. Effects of captopril against radiation injuries in the Gottingen minipig model of hematopoietic-acute radiation syndrome. *PloS One.* 2021;16:e0256208.
- [152] Arnett DK, Khera A, Blumenthal RS. 2019 ACC/AHA guideline on the primary prevention of cardiovascular disease: part 1, lifestyle and behavioral factors. *JAMA Cardiol.* 2019;4:1043-4.

- [153] Cleveland Clinic. Statin Medications & Heart Disease. 2019. Available at: <https://my.clevelandclinic.org/health/articles/17506-statin-medications--heart-disease> [Last accessed December 15, 2021]
- [154] Camara Planek MI, Silver AJ, Volgman AS, Okwuosa TM. Exploratory review of the role of statins, colchicine, and aspirin for the prevention of radiation-associated cardiovascular disease and mortality. *J Am Heart Assoc.* 2020;9:e014668.
- [155] Efimova EV, Ricco N, Labay E, Mauceri HJ, Flor AC, Ramamurthy A, et al. HMG-CoA reductase inhibition delays DNA repair and promotes senescence after tumor irradiation. *Mol Cancer Ther.* 2018;17:407-18.
- [156] Graaf MR, Beiderbeck AB, Egberts AC, Richel DJ, Guchelaar HJ. The risk of cancer in users of statins. *J Clin Oncol.* 2004;22:2388-94.
- [157] Nielsen SF, Nordestgaard BG, Bojesen SE. Statin use and reduced cancer-related mortality. *New Engl J Med.* 2012;367:1792-802.
- [158] Alexandre L, Clark AB, Bhutta HY, Chan SS, Lewis MP, Hart AR. Association between statin use after diagnosis of esophageal cancer and survival: A population-based cohort study. *Gastroenterology.* 2016;150:854-65 e1; quiz e16-7.
- [159] Ziegler V, Henninger C, Simiantonakis I, Buchholzer M, Ahmadian MR, Budach W, et al. Rho inhibition by lovastatin affects apoptosis and DSB repair of primary human lung cells in vitro and lung tissue in vivo following fractionated irradiation. *Cell Death Dis.* 2017;8:e2978.
- [160] Chung KJ, Park KR, Lee JH, Kim TG, Kim YH. Simvastatin reduces capsular fibrosis around silicone implants. *J Korean Med Sci.* 2016;31:1273-8.
- [161] Fritz G, Henninger C, Huelsenbeck J. Potential use of HMG-CoA reductase inhibitors (statins) as radioprotective agents. *Br Med Bull.* 2011;97:17-26.
- [162] Ma S, Ma CC. Recent development in pleiotropic effects of statins on cardiovascular disease through regulation of transforming growth factor-beta superfamily. *Cytokine Growth Factor Rev.* 2011;22:167-75.
- [163] Williams JP, Hernady E, Johnston CJ, Reed CM, Fenton B, Okunieff P, et al. Effect of administration of lovastatin on the development of late pulmonary effects after whole-lung irradiation in a murine model. *Radiat Res.* 2004;161:560-7.
- [164] Ostrau C, Hulsbeck J, Herzog M, Schad A, Torzewski M, Lackner KJ, et al. Lovastatin attenuates ionizing radiation-induced normal tissue damage in vivo. *Radiother Oncol.* 2009;92:492-9.
- [165] Zhao X, Yang H, Jiang G, Ni M, Deng Y, Cai J, et al. Simvastatin attenuates radiation-induced tissue damage in mice. *J Radiat Res.* 2014;55:257-64.

- [166] Doi H, Matsumoto S, Odawara S, Shikata T, Kitajima K, Tanooka M, et al. Pravastatin reduces radiation-induced damage in normal tissues. *Exp Ther Med.* 2017;13:1765-72.
- [167] Haydont V, Bourcier C, Pocard M, Lusinchi A, Aigueperse J, Mathe D, et al. Pravastatin Inhibits the Rho/CCN2/extracellular matrix cascade in human fibrosis explants and improves radiation-induced intestinal fibrosis in rats. *Clin Cancer Res.* 2007;13:5331-40.
- [168] Singh VK, Seed TM. A review of radiation countermeasures focusing on injury-specific medicinals and regulatory approval status: part I. Radiation sub-syndromes, animal models and FDA-approved countermeasures. *Int J Radiat Biol.* 2017;93:851-69.
- [169] Sun H, Tsai Y, Nowak I, Liesveld J, Chen Y. Eltrombopag, a thrombopoietin receptor agonist, enhances human umbilical cord blood hematopoietic stem/primitive progenitor cell expansion and promotes multi-lineage hematopoiesis. *Stem Cell Res.* 2012;9:77-86.
- [170] National Aeronautics and Space Administration. NASA technology transfer program: Dietary formulas fortify antioxidant supplements. 2012. Available at: [https://spinoff.nasa.gov/Spinoff2012/hm\\_2.html](https://spinoff.nasa.gov/Spinoff2012/hm_2.html) [Last accessed December 17, 2021]
- [171] Hirouchi T, Ito K, Nakano M, Monzen S, Yoshino H, Chiba M, et al. Mitigative effects of a combination of multiple pharmaceutical drugs on the survival of mice exposed to lethal ionizing radiation. *Curr Pharm Biotechnol.* 2015;17:190-9.
- [172] Patchen ML, Fischer R, MacVittie TJ. Effects of combined administration of interleukin-6 and granulocyte colony-stimulating factor on recovery from radiation-induced hemopoietic aplasia. *Exp Hematol.* 1993;21:338-44.
- [173] Day RM, Davis TA, Barshishat-Kupper M, McCart EA, Tipton AJ, Landauer MR. Enhanced hematopoietic protection from radiation by the combination of genistein and captopril. *Int Immunopharmacol.* 2013;15:348-56.
- [174] Patchen ML, MacVittie TJ, Solberg BD, D'Alesandro MM, Brook I. Radioprotection by polysaccharides alone and in combination with aminothiols. *Adv Space Res.* 1992;12:233-48.
- [175] Patchen ML, MacVittie TJ, Souza LM. Post-irradiation treatment with granulocyte colony-stimulating factor and preirradiation WR-2721 administration synergize to enhance hemopoietic reconstitution and increase survival. *Int J Radiat Oncol Biol Phys.* 1992;22:773-9.
- [176] Smith SM, Zwartz SR, Heer M. Human adaptation to space flight: The role of nutrition. 2014. Available at: <https://www.nasa.gov/sites/default/files/human-adaptation-to-spaceflight-the-role-of-nutrition.pdf> [Last accessed March 1, 2022]
- [177] National Aeronautics and Space Administration. Food and nutrition - Technical brief. 2022. Available at: [https://www.nasa.gov/sites/default/files/atoms/files/food\\_and\\_nutrition\\_technical\\_brief\\_ochmo\\_06052020\\_rev\\_a.pdf](https://www.nasa.gov/sites/default/files/atoms/files/food_and_nutrition_technical_brief_ochmo_06052020_rev_a.pdf) [Last accessed March 1, 2022]

[178] National Aeronautics and Space Administration. <https://science.nasa.gov/science-news/news-articles/a-successful-mission-starts-with-nutrition>. 2019. Available at: <https://science.nasa.gov/science-news/news-articles/a-successful-mission-starts-with-nutrition>  
[Last accessed March 1, 2022]

## Countermeasures for extraterrestrial radiation exposures and related injuries



Singh VK, Seed TM. MCMs for extraterrestrial environments

Figure 1

**Table 1.** Current radiation countering medicinals within space vehicle’s emergency medical kit

| #      | Drug type/<br>class                   | Mechanism of<br>action  | Intended use   | Drug name   | Drug maker | Route/ dose                                |
|--------|---------------------------------------|---|--|---|------------|--|
| 1<br>a | Anti-emetics                          | 5-HT3 receptor antagonist   | Prevent nausea/vomiting  | Ondansetron (Zofran) (or Gransietron - Kytril)  | Generic    | Oral/1 tablet/d (8 – 24 mg)                |
| 1<br>b | Anti-inflammatories                   | Glucocorticoid receptor agonist                                   | Minimize inflammation associated pain  | Dexamethosone (Decadron®)   | Generic    | Oral-liquid or tablet – 1 d                |
| 1<br>c | Anti-diarrheals                       | Opioid $\mu$ receptor agonist                                     | Diarrhea   | Loperamide hydrochloride Imodium®   | Generic    | Oral/1 – 2 tablets (2 mg/tab); as required |
| 2<br>a | Antimicrobials - fluoroquinolone      | Inhibits bacterial DNA replication                                | Broad spectrum-prevention/control of mixed infections - Gram negative microbes   | Ciprofloxacin   | Generic    | Oral or injection 250 – 500 mg/12 h/d      |
| 2<br>b | Antimicrobials - beta lactams         | Inhibits mainly gram positive microbes                            | Inhibit microbial cell wall synthesis  | Cephalosporin Cephradine or Ceftoaroline 1 <sup>st</sup> & 5 <sup>th</sup> generation | Generic    | Orally or injection                        |
| 2<br>c | Antimicrobials - beta lactams         | Inhibits gram positive microbes                                   | Inhibits microbial cell wall synthesis   | Penicillin e.g., penicillin V potassium   | Generic    | Oral tablets/250 mg/tablet                 |
| 2<br>d | Antimicrobials - macrolide            | Inhibits gram positive microbes, with wider range than penicillin | Inhibits microbial protein synthesis via binding to ribosomal binding & limiting | Macroclides e.g., erythromycin, clarithromycin, azithromycin                          | Generic    | Oral tablets/333 mg/8 h                    |
| 3<br>a | Skin treatments - Sulfa drugs         | Bactericidal  | radiation burns & blisters anti-infectives                                       | Silver sulfadiazine Silvadene   | Generic    | Topical                                    |
| 3<br>b | Skin treatments - anti-inflammatories | Suppression of immunologic mediators of inflammation              | Irradiation blisters/burns anti-inflammatory                                     | Corticosteroids   | Generic    | Topical                                    |
| 3<br>c | Skin treatments - analgesics          | Suppression of pain via neutral mediators                         | radiation blisters/burns analgesic   | Opioids   | Generic    | Topical/oral                               |
| 3<br>d | Skin treatments - topicals            | Control of skin barrier damage                                    | irradiation blisters/burns lesion treatment(s)                                   | Crystalloid solutions Sterile gauze   | Generic    | Topical                                    |
| 4      | Dehydration                           | Cutaneous water loss  | radiation associated dehydration   | <i>iv</i> provided physiological saline solutions                                     | Generic    | <i>iv</i> infusions                        |

Inclusion of the hematopoietic growth factor, rhuG-CSF, is under consideration.  
5-HT3, 5-hydroxytryptamine (serotonin)

**Table 2.** Select recombinant growth factors: Possible candidates for inclusion in the space radiation medical emergency kit

| # | Drug class                    | Intended use  | Drug name                    | Manufacturer   | Dosage/dosing   |
|---|-------------------------------|---|------------------------------|----------------|---|
| 1 | Recombinant GF<br>rhuG-CSF    | Mitigate acute granulocytopenia<br>Stimulate granulocytopenia                                     | Filgrastim -<br>Neupogen     | Amgen          | Injection - 10 µg/kg/d<br>Initiate - 1 d post irradiation<br>Multiple injections until ANC's<br>> 1,000/µl for 3 consec d                                       |
| 2 | Recombinant GF<br>rhuPEGG-CSF | Mitigate acute granulocytopenia<br>Stimulate granulocytopenia                                     | PEG-filgrastim -<br>Neulasta | Amgen          | Injection - 6 mg x 2<br>Initiate - 1 d post-irradiation<br>2 injections at 1 & 8 d  |
| 3 | Recombinant GF<br>rhuGM-CSF   | Mitigate granulo/monocytopenia<br>Stimulate granulo/monocytopenia                                 | Sargogastim -<br>Leukine     | Sanofi-Aventis | Injection - 7 µg/kg/d<br>Initiate - 2 d post-irradiation<br>Multiple injections   |
| 4 | GF receptor agonist<br>TPO    | Mitigate thrombocytopenia, stimulate<br>thrombocytopenia  | Romboplastim -<br>Nplate     | Amgen          | Injection - 10 µg/kg once<br>Initiate - 1 d post-irradiation<br>Single injection  |
| 5 | Recombinant GF<br>rhuEPO      | Mitigate anemia, stimulate<br>erythropoiesis  | EPO - Epogen                 | Amgen          | Injection/infusion - 150 U/kg/3x<br>weekly<br>Initiate- post-irradiation with<br>presentation of severe anemia<br>(e.g., Hg < 9-10 g/dl)<br>Multiple injections |
| 6 | Recombinant GF<br>rhuKGF      | Prevent/mitigate chemo/radio-<br>pathology of epithelium, stimulate<br>epithelial repair/regrowth | Palifermin -<br>Kepivance    | Amgen          | Infusions - 60 µg/kg/6x, multiple<br>infusions - 3 prior, plus 3<br>following exposure to cytotoxic<br>agent.   |

**Table 3.** A representative sampling of reengineered radioprotective drugs for possible use within the extraterrestrial space environment

| # | Drug class                 | Drug name                         | Modification                        | Intended use  | Manufacturer  |
|---|----------------------------|-----------------------------------|-------------------------------------|---|---|
| 1 | Recombinant interleukin    | BBT-059 - PEG-IL-11               | PEGylation                          | Mitigation-acute radiation-induced GIS-ARS  | Boulder Pharmaceuticals   |
| 2 | Nutraceutical              | BIO 300                           | Nanoparticulization via Wet-milling | Prevention/mitigation – radiation injury  | Neumedicines  |
| 3 | Toll like receptor agonist | Entolmolid - CBLB502              | Recombinant molecular reengineering | Prevention/mitigation – radiation-induced acute hematopoietic injury                                | Cleveland BioLabs   |
| 4 | Aminothiols                | PEG-amifostine - encap-amifostine | PEGylation, encapsulation           | Systemic radioprotection /mitigation of acute, early and delayed or late-arising radiation injuries | None - reported by research institutes, Chinese Mil Institute, AFRI |

**Table 4.** A partial list of commonly used and marketed pharmaceuticals with documented radioprotective attributes and with potential to be repurposed for the space environment

| <b>Drug class</b>               | <b>Generic name</b>                    | <b>Primary use</b>   | <b>Repurposed use</b>   | <b>Delivery route/<br/>Dosing<sup>5</sup></b>   |
|---------------------------------|--|--|---|---|
| <b>1</b> ACE inhibitors         | e.g., Captopril (~1/9 alike drugs)     | Hypertension/cardiovascular disease  | Mitigate acute/chronic radiation injuries of blood/marrow, heart & vasculature, lung & kidney   | Oral/daily dosing/TBD (e.g., 10 mg/d each day as needed)  |
| <b>2</b> Statins                | e.g., Atorvastatin (~1/ 'alike' drugs) | Regulate cholesterol metabolism/mitigate hypertension & cardiovascular disease                             | Mitigate- acute/chronic radiation injuries- blood/marrow, gastrointestinal & heart, mitigate radiation cancer risks   | Oral/daily dosing/TBD (e.g., 20 mg/d each day as needed)  |
| <b>3</b> Anti-inflammatories    | NSAIDs e.g., Acetaminophen             | Reduce inflammation/reduce pain  | Reduce radiation associated inflammation/cytokine storm   | Oral, injection, or topical dosing/TBD (e.g., oral dosing - 500 mg/6 – 12 h each day as needed) |
| <b>4</b> Amino thiols           | NAC e.g., NAC                          | Nutritional supplement - anti-oxidant/detoxifier /glutathione regulator/ alleviates pulmonary inflammation | Prevent/mitigate acute radiation associated injuries via free radical quenching, possible mitigation of radiation associated cancer risks   | Oral dosing/TBD (e.g., oral daily dosing of 600 – 1200 mg/d)                                    |
| <b>5</b> Hemorheological agents | Pentoxifylline                         | Improves arterial blood flow/ improves tissue oxygenation / alleviates muscle cramps                       | Prevent acute radiation injuries of select vital organ systems; pulmonary, vascular, hematopoietic and gastrointestinal via anti-oxidant, anti-inflammatory, immune-modulating effects with improved blood flow | Oral dosing/TBD (e.g., oral dosing of ~400 mg/kg)   |
| <b>6</b> Nutraceuticals         | Tocotrienols e.g., gamma tocotrienol   | Nutritional supplement- antioxidant/anti-inflammatory effects  | Prevent/mitigate acute radiation associated injuries via anti-inflammatory & anti-oxidant properties  | Oral or injection dosing/TBD (e.g., Oral dosing - 50 mg/tablet/d)                               |
| <b>7</b> Nutraceuticals         | Genistein                              | Nutritional supplement-immune support/ bone health   | Prevent/mitigate acute radiation associated injuries via free radical quenching. Mitigate long-term radiation cancer risks  | Oral or injection dosing/TBD (e.g., oral dosing)  |
| <b>8</b> Nutraceuticals         | Melatonin <sup>6</sup>                 | Nutritional supplement-immune system support /sleep aide   | Prevent/mitigate acute radiation associated injuries, immune system support/sleep aide  | Oral dosing/TBD (e.g., oral dose of 1 – 5 mg/tablet/ dose/d as needed)                          |

Listed dosing/dosing regimens are all based on those commonly used terrestrially: extraterrestrial dosing levels that are both safe and effective will need to be determined.

**Table 5.** A selection of potentially useful drug combinations that might serve as effective radiation countermeasures for the extraterrestrial environment

| #  | Drug class combination                              | 1 <sup>st</sup> drug- MCM effect/target         | 2 <sup>nd</sup> drug- MCM effect/target   | Expected clinical outcome/advantage  |
|----|---|---|---|--|
| 1  | Granulopoietic GF plus thrombopoietic GF            | PEGfilgrastim sparing hematopoietic function(s) | Romiplostim - Sparing hematopoietic function(s)   | Enhanced recovery from acute radiation-induced ( <i>SPEs</i> ) granulocytopenia and thrombocytopenia, with enhanced survival   |
| 2  | Granulopoietic GF and/or granulomonopoietic GF      | PEGfilgrastim Sparing hematopoietic function(s) | Sargramostim - Sparing hematopoietic function(s)  | Enhanced recovery from acute radiation-induced ( <i>SPEs</i> ) granulocytopenia, monocytopenia, and thrombocytopenia with enhance survival   |
| 3  | Granulopoietic GF plus thrombopoietic GF            | PEGfilgrastim Sparing hematopoietic function(s) | IL-11/Neumega - Sparing marrow & GI functions   | Enhanced marrow and gastrointestinal tissue recovery from acute radiation-induced ( <i>SPEs</i> )  |
| 4  | ACEi plus recombinant GF                            | Captopril (or alike ACEi)                       | Romiplostim (or alike - e.g., <i>Eltrombopag</i> <sup>1</sup> )   | Enhanced protection/recovery of hematopoietic tissues from acute irradiation ( <i>SPEs</i> ) exposures and chronic GCR exposures   |
| 5  | ACEi plus radioprotective nutraceutical             | Captopril (or alike ACEi)                       | BIO300 genistein  | Enhanced protection/recovery of hematopoietic tissues from acute radiation ( <i>SPEs</i> ) exposures and chronic GCR exposures   |
| 6  | ACEi plus radioprotective aminothioliol             | Captopril (or alike ACEi)                       | Amifostine (or alike aminothioliol)   | Mitigation of space radiation-elicited acute/chronic morbidities (cancers, tissues fibrosis) and associated mortality risks.   |
| 7  | ACEi plus statin                                    | Captopril (or alike ACEi)                       | Atorvastatin  | Mitigation of space radiation-elicited chronic morbidities (e.g., cancers; tissue fibrosis) and reduced mortality risks.   |
| 8  | Tocol nutraceutical plus pentoxifylline             | Gamma tocotrienol (or alike tocol)              | Pentoxymfylline (or alike phosphordiesterase inhibitor)   | Enhanced radioprotective/ radiomitigating actions of tocol nutraceutical. Ameliorates radiation-associate vascular and gastrointestinal injuries   |
| 9  | Tocol nutraceutical plus ACEi                       | Gamma tocotrienol (or alike tocol)              | Captopril (or alike ACEi)   | Extended, enhanced radioprotection of coverage. Vital organ systems (marrow, gut, lung, kidney) against both acute ( <i>SPEs</i> ) and chronic ( <i>GCRs</i> ) space irradiation associated syndromes  |
| 10 | Tocol nutraceutical plus benzylsulfone              | Gamma tocotrienol (or alike tocol)              | Recilisib (ExRad) <sup>2</sup><br>Chlorobenzylsulfone   | Promotes radioresistance of hematopoietic and gastrointestinal tissues and limits long-terms pathologically risks of ionizing radiation exposures.   |
| 11 | Tocol nutraceutical plus statin                     | Gamma tocotrienol (or alike tocol)              | Atvorastatin (or alike statin)  | Promotes protection and function of vital organ systems. Limits 'late-effects' of radiation exposures (cancers) & associated mortality risks.  |
| 12 | Gene/gene product transfer plus dietary supplements | MnSOD plasmid/liposome gene transfer            | Antioxidant dietary supplements (e.g., blueberry/fruit extracts or NASA /AmeriScience dietary supplement <sup>3</sup> ) | Protection/mitigation of both intense ( <i>SPEs</i> ) space ionizing radiation exposures and associated acute morbidities, as well as chronic ( <i>GCRs</i> ) ionizing radiation exposures that elicit late arising pathologies (e.g., tissue fibrosis, cancer, behavioral and cognitive dysfunctions) |